Increase in Bcl₂ expression of penile and prostate cells of Sprague Dawley male rats following treatment with buceng (combination of *Pimpinella alpina* molk with *Eurycoma longifolia* Jack)

Taufiqurrachman Nasihun

Department of Biochemistry and Andrology, Faculty of Medicine, Sultan Agung Islamic University, Semarang, Indonesia

ABSTRAK

Latar belakang: Pemberian buceng kombinasi pasak bumi (Eurycoma longifolia Jack) dan purwoceng (Pimpinella alpine Molk) telah terbukti mampu meningkatkan kadar testosteron, menurunkan apoptosis, dan ekspresi caspase₃. Bcl₂ adalah protein anti apoptosis yang ditemukan dalam sitoplasma. Tujuan penelitian ini adalah untuk mengungkap efek buceng terhadap ekspresi Bcl₂ pada jaringan penis dan prostat tikus.

Metode: Studi experimental dengan 24 tikus jantan Sprague Dawley umur 90 hari, berat badan \pm 300 gram, dibagi menjadi 4 kelompok secara acak. Kelompok A tikus normal, kelompok B tikus dikastrasi dan diberi buceng 100 mg/hari, per oral (Cast-Bcg). Kelompok C tikus dikastrasi dan diberi air 2 ml sebagai plasebo dari buceng (Cast-Plac). Kelompok D tikus dikastrasi, diberi mesterolon 6,75 mg/hari, per oral sebagai testosteron eksogen (Cast-Mest). Pengamatan dilakukan selama 30 hari. Uji Mannova digunakan untuk menguji perbedaan ekspresi Bcl₂ di antara kelompok dengan tingkat signifikansi $p \leq 0.05$.

Hasil: Proses kastrasi menurunkan ekspresi Bcl_2 pada penis dan prostat (53,0 dan 50,9%) secara signifikan dibanding tikus normal (82,6 dan 84,2%, p < 0.001). Pemberian mesterolon menyebabkan peningkatan ekspresi Bcl_2 (77,1 dan 78,1%) mendekati normal. Normalisasi ekspresi Bcl_2 juga diperoleh dengan pemberian buceng (73,8 dan 782%).

Kesimpulan: Pemberian buceng dapat meningkatkan ekspresi Bcl₂ pada jaringan penis dan prostat, setara dengan normal pada tikus kastrasi yang mendapat mesterolon.

ABSTRACT

Background: Treatment with *buceng* combination of *Eurycoma longifolia* Jack and *Pimpinella alpine* Molk has been proven to increase testosterone level, decrease apoptosis and caspase₃ expression. Bcl₂ is an antiapoptotic protein found in cytoplasm which inhibits cells apoptosis. This study was aimed to investigate the effect of buceng on Bcl₂ expression on penile and prostate tissues of the rats.

Methods: In this experimental study, 24 male Sprague Dawley rats of 90 days old, weighing \pm 300 grams, were randomly assigned into four groups. Group A, normal rats. Group B, castrated rats and treated with *buceng* 100 mg/day, per oral (Cast-Bcg); Group C, castrated rats and treated with 2 ml of water as placebo against buceng (Cast-Plac). Group D, castrated rats, treated with mesterolone 6.75 mg/day, per oral, as exogenous testosterone (Cast-Mest). All rats were treated for 30 days. Manova test was used to analyze the different expression of Bcl, among groups with significance level at p \leq 0.05.

Results: Castration was associated with significant decrease of Bcl₂ expression in the penile and prostate tissues (53.0 and 50.9%, respectively) compared to normal rats (82.6 and 84.2%, respectively, p < 0.001). Treatment with mesterolone reversed Bcl₂ expression (77.1 and 78.1%) to a near normal level. The same level of Bcl₂ expression was also observed with *buceng* treatment (73.8 and 78.2%).

Conclusion: The treatment with *buceng* could enhance Bcl₂ expression in penile and prostate tissues, comparable to normal rats and mesterolone treated rats.

Keywords: apoptosis, Bcl₂, *buceng*, testosterone

pISSN: 0853-1773 • eISSN: 2252-8083 • http://dx.doi.org/10.13181/mji.v24i1.1023 • Med J Indones. 2015;24:8-13 • Received 29 Aug 2014 • Accepted 26 Feb 2015

Correspondence author: Taufigurrachman Nasihun, taufig_rn@yahoo.com

Copyright @ 2015 Author. This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License (http://creativecommons.org/licenses/by-nc-sa/4.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original author and source are properly cited.

Pasak bumi (Eurycoma longifolia Jack) and purwoceng (Pimpunella alpina Molk) both are Indonesian indigenous plants^{1,2} combination of them so called buceng. Treatment of buceng extract has been proven to increase serum testosterone level, suppress apoptosis, and decrease caspase, expression.³⁻⁵ There are growing evidences that apoptosis process has strong correlation with decrease in antiapoptotic protein, on one hand, and on the other hand increase in proapoptotic protein.⁶ Caspase, is a proapoptotic protein that serves as an executor of apoptosis,⁷ whereas Bcl₂ is an antiapoptotic protein preventing apoptosis.8 Based on these facts, the decrease in both apoptosis and caspase, in buceng treatment, physiobiologically will be followed by increase in Bcl₂ expression. However, the effect of buceng extract treatment on Bcl, expression is still unknown.

The influence of buceng on Bcl₂ expression is important to investigate, since many antiapoptotic proteins regulate the apoptosis process. When buceng is proven capable to increase in Bcl, expression, will clarify the mechanism by which buceng reduces apoptosis. To date, studies on buceng have been extended to the increase of testosterone, luteinizing hormone (LH), follicle stimulating hormone (FSH), decrease in apoptosis and caspase₃,³⁻⁵ whereas the effect of buceng on Bcl₂ expression has not been elucidated yet. Decrease in apoptosis pathway after buceng treatment is necessary to be explored, since the apoptosis process can be regulated by either caspase, or Bcl₂ and by both caspase, and Bcl₂.^{7,8} In line with those arguments, study on buceng effect on Bcl₂ expression and its role in the downregulation of apoptosis in order to enhance male vitality is necessary.

Bcl₂ is an anti apoptosis protein, existing in cell cytoplasm and has role in preventing the release of Cyt C and diablo/Smac, a protein that binds inhibitor apoptosis protein (IAP) from mitochondria.^{9,10} When released into cytoplasm, diablo/Smac will bind IAP in order to activate apoptosome and caspase and then induce cell to undergo apoptosis.^{9,10} In addition, Bcl₂ is an antiapoptototic protein whose activity is dependent on testosterone level.¹¹ Buceng, has previously been proven to up-regulate Te, LH, FSH level, down-regulate apoptosis process and caspase₂ expression in Sprague Dawley male rats.^{4,5} However, its effect on Bcl₂ expression has not yet been explored. Thus, the aim of the present study was to elucidate the effect of buceng on Bcl₂ expression in penile and prostate tissues of Sprague Dawley male rats.

METHODS

This was an experimental study on 24 Sprague Dawley male rats, 90 days old, and body weight of 300 grams. All animals were randomly allocated into 4 groups, consisting of 6 rats in each group. Group A, normal rats. Group B, castrated rats and treated with buceng of 100 mg/day in 2 ml water, per oral (Cast-Bcg). Group C, castrated rats and treated with 2 ml of water as placebo against buceng (Cast-Plac). Group D, castrated rats, treated with mesterolone 6.75 mg/day, per oral, as exogenous testosterone (Cast-Mest). All rats were treated for 30 days.

Buceng extract was obtained from soxhlet extraction method with water as a solvent. The effective dose of buceng 100 mg/day has been determined according to the buceng previous study.^{4,5} All rats underwent acclimatization for a week in individual cage. All rats got daily chow and ad libitum access to tap water. After a week acclimatization, rats in group A were sacrificed and taken their tissues of penis and prostate for Bcl₂ expression examination as normal reference. Rats were given buceng in group B, aquadest in group C, mesterolone in group D orally for 30 days at 7 a.m. After treated for consecutive 30 days, rats in group B, C, and D were sacrificed and their penile and prostate tissues taken for measurement of Bcl₂ expression. Measurements of Bcl expression were performed bv immunohistochemical method (kit: Bcl-2/100/ D5 from novocastra) and then observed by light microscope in 400x magnification in each 5 fields. The Bcl₂ expression was calculated as percentage of total Bcl₂ expressing cells to total cells (total Bcl₂ expressing cells/total cells x 100). Before examination, all penile and prostate tissues were frozen in nitrogen solution and kept in temperature of -20°C.12

The experimental study has taken place at the Animal Experimental Development Unit and Pathological Anatomy Sardjito Hospital/School of Medicine Gadjah Mada University. Manova test was used to examine the difference among groups, and HSD Tukey was implemented to evaluate the difference among groups. The significant level of all examinations was set at $p \le 0.05$.

RESULTS

Genetic and age variability of rats in the present study can be neglected, since all rats were bred in the same growth condition. The 30 days treatment resulting Bcl_2 expression both in penile and prostate, shown in table 1.

As shown in table 1 both Bcl_2 expression in penis and prostate were significantly different among 4 groups, p < 0.001.

Difference of Bcl₂ expression of penile

Observation with 400x magnification of light microscope on immunohistochemical stainning of penile tissues can be seen in figure 1.

The numbers of cells (cytoplasm), in which Bcl_2 positive expression (brown) and Bcl_2 negative expression (blue) in each five observation fields were counted as shown in table 1. That table indicates that the highest percentage mean of Bcl_2 positive expression was in normal group,

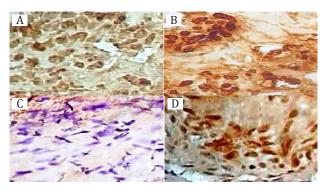


Figure 1. Bcl₂ expression in penile tissue. a) normal, b) castrated-buceng group, c) castrated-placebo group, d) castrated-mesterolone group.

followed by Cast-Bcg group, Cast-Mest group, and the lowest was in Cast-Plac group (figure 2).

Difference of Bcl₂ expression of prostate

Light microscopic scanning with 400x magnification on immunohistochemically stained prostate tissues preparation provides results as shown in figure 3.

All data were collected from these observation, and the numbers of cells (cytoplasm) in which Bcl₂ expression was positive (brown) and Bcl₂

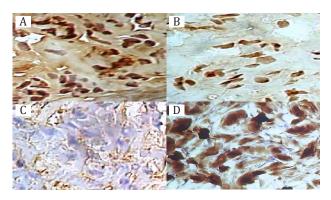


Figure 2. Bcl₂ expression in prostat tissue. a) normal, b) castrated-buceng group, c) castrated-placebo group, d) castrated-mesterolone group

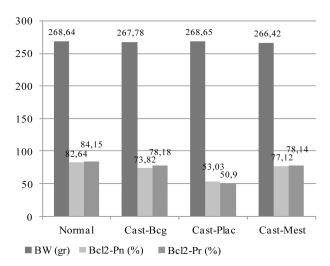


Figure 3. Body weight and Bcl_{2} expression in penile and prostate cells

Table 1	. Bod	y weig	ht and	Bcl ₂	expressi	on in	penile	and	prostate	cells	
		5 - 0		- 2	- I		1 .		I		

_	Groups							
Variables	A (Normal) (n = 6)	B (Cast-Bcg) (n = 6)	C (Cast-Plac) (n = 6)	D (Cast-Mest) (n = 6)	p (Manova)			
BW (gram ± SD)	268.64 ± 28.03	267.78 ± 25.20	268.65 ± 28.53	266.42 ± 19.55	0.994			
Bcl_2 penile (% ± SD)	82.64 ± 3.70	73.82 ± 8.40	53.03 ± 21.3	77.12 ± 6.20	0.000			
Bcl ₂ prostate (% ± SD)	84.15 ± 4.07	78.18 ± 7.83	50.90 ± 21.84	78.14 ± 5.40	0.000			

expression was negative (blue) from each five fields observation were counted and results are shown in table 1. This table indicates that the highest Bcl₂ positive expression was in normal group, followed by Cast-Bcg group, Cast-Mest group, and the lowest was in Cast-Plac group (figure 3).

DISCUSSION

The results of the present study indicate that buceng is capable to increase Bcl₂ expression of penile and prostate cells comparable to that of mesterolone and endogenous testosterone. Thus, the effect of buceng is attributable to up-regulation of Te level. This statement is in accordance with previous studies showing that buceng has been proven capable to up-regulate testosterone level in Sprague Dawley male rats as well as able to decrease caspase, expression and in turn apoptosis of penile and prostate cells.^{4,5} Caspase₃ is an IL-1 beta converting enzyme (ICE) like protease that is able to break peptide chains at the point following aspartic acid residues.¹³ At least, there are 10 types of caspases that have been identified and engaged in apoptosis process. Many of those caspases were arranged in a hierarchy, beginning from caspase, $caspase_{8}$, $caspase_{9}$, and $caspase_{10}$ as initiator caspase, then activates caspase, which serves as an amplifier,^{14,15} and finally activates caspase, caspase, and caspase, as an execution machinery. Of those various caspases are inhibited by Bcl₂, a protein of cytoplasm that extremely testosterone dependent.11 In conjunction with Bcl_{v1} they constitute antiapoptotic protein.¹⁴ An important result of this present study is that the increase in Bcl₂ expression of penile and prostate cells in response to the treatment with buceng extract which commensurates with normal and mesterolone groups.

Mesterolone (17-betahydroxy-1-alpha-methyl-5alpha-androstan-3-one, $C_2OH_32O_2$), is a derivative of dehidrotestosteron (DHT) that is frequently used as replacement therapy in male hypogonadism.¹⁶ It can be administered orally and rapidly absorbed without metabolism or inactivation in the liver. Thus, testosterone serum level will increase in accordance with the dose given.¹⁷ In this study, the dose of mesterolone was 6.75 mg/day or equivalent to 0.02 - 0.06 mg/kg BW/day, based on the physiological calculation for 70 kg man.¹⁸ Higher or supraphysiologic dose of mesterolone will result in intoxication indicated by weak, sluggish, even unmoved rats.

Compared to Protodiocin (Tribullus terestris L), a natural androgen that has long been launched in the market,¹⁹ buceng that also has long been utilized to increase male vitality and sexual function since about 3 centuries ago posseses a little bit superiority in improving sexual and reproductive potencies in male hypogonadism. Various evidences indicated that protodiocin was capable to improve sexual behavior parameters such as intra-cavernous pressure, testosterone level, DHT, DHEA, and NO level in animal models.²⁰ Likewise, buceng was capable to increase testosterone level, endothelial nitric oxide synthase (eNOS) and neuronal nitric oxide synthase (nNOS), and cyclic guanosin monophosphate (cGMP).²¹ Moreover, buceng was able to stimulate both LH and FSH secretions,³ whereas protodioscin just stimulates LH secretion without affecting increased FSH secretion.^{22,23} Increase in LH secretion results in increasing testosterone level secreted by Leydig cells in testis, while increased FSH secretion induces LH receptor expression on Leydig cells, and in turn up-regulates testosterone level, subsequently increases sexual function and spermatogenesis.²⁴ As such, an illustration may be presented a report from Phillip and colleagues in 1998 that the patient who had the defect in the beta subunit of FSH and FSH receptor will show delayed puberty and had low serum testosterone and FSH but high LH concentrations.²⁵ Albeit that report was not exactly appropriate, nonetheless it indicates that the existence of both LH and FSH are essential for maintenance of testicular function like buceng is. The probable similarity in effect of protodiocin and buceng is attributable to the similarity in chemical structure of furostanol contained in protodiocin and sitosterol in buceng. The similarity of chemical structure lies on OH group bound to carbon atom 3 in the nucleus of cyclopentanoprehidrofenantren. The OH group leads to formation of glycoside binding between sitosterol (aglicon) and olygosacharide (hexose or mannose). Consequently, sterol physical properties might be transformed from nonpolar to polar ones.²⁶ Based on the effect and structure similarities of furostanol and buceng, deduction can be proposed that sitosterol of buceng is included in the family of furostanol compound which is capable to increase DHEA level as well as testosterone.²⁰ Accordingly, both buceng and furostanol can be used as hormone replacement therapy in male hypogonadism.

Male hypogonadism is associated with significant increase of morbidity and decline in quality of life, especially related to skeletal muscles frailty caused by sarcopenia, and sarcopenia due to apoptosis.^{16,27} Various evidences indicated that high level of testosterone capable to decrease in apoptosis on any cells that possess testosterone receptors.^{11,28} On the other hand, down-regulation of testosterone level gives rise various organs, in particular penile and prostate which is regulated by testosterone experiencing reduction of size and function mediated by increasing apoptosis.¹¹ The previous study showed that buceng was capable to increase testosterone level,³ and in the present study indicates that buceng is capable to increase Bcl₂ expression in penile and prostate tissues, hence decrease their apoptosis.⁴ Taken together, the results of the previous and the present studies and in line with biological plausibility, could explain the pathway of buceng effect on apoptosis in penile and prostate tissues. The explanation of that pathway is that buceng could inhibit apoptosis through increasing testosterone level and Bcl₂ expression, followed by decreasing caspase, expression, and then apoptosis. It can be expected that buceng can be used as a hormone replacement therapy related to male hypogonadism or infertility. In addition, buceng is relatively safe, since the majority of males who experienced decrease in vitality have used it since 3 centuries ago, without adverse effect that has been reported.

In conclusion, the treatment of buceng with dose of 100 mg/day for consecutive 30 days in castrated Sprague Dawley male rats is capable to increase in Bcl₂ of penile and prostate cells, comparable to that in normal rats and mesterolone treated castrated rats.

Acknowledgment

The author greatly appreciates Prof. Dr. Susilo Wibowo, MS. Med Sp.And; Prof. Dr. Suhartono Taat Putra; Prof. Tjahjono, Sp.PA; Prof. Sisminadri, Apt.; Dr. Haryadi, and Ms. Yuli as a laborant who assisted in carrying out this study.

Conflict of interest

The author affirm no conflict of interest in this study.

REFERENCES

- 1. Nainggolan O, Simanjuntak JW. Pengaruh ekstrak etanol akar pasak bumi (*Eurycoma longifolia* Jack) terhadap perilaku seksual mencit putih [the effect of *Eurycoma longifolia* Jack ethanol extract on sexual behaviour in mice]. Cermin D Kedokteran. 2005;146:55-7. Indonesian.
- 2. Darwati I, Roostika I. The *Pimpinella alpina* Molk research status in Indonesia. Buletin P Nutfah. 2006;12(1):9-15. Indonesian.
- 3. Taufiqurrachman. The effect of buceng extracts on androgen production in Sprague Dawley male rats. Med J Indones. 2012;21(1):28-31.
- 4. Taufiqurrachman. Extract of *Pimpinella alpina* Molk and *Eurycoma longifolia* Jack increase testosterone level, decrease apoptosis in penis and prostate cells. Med M Indonesiana. 2012;46:34-40.
- Taufiqurrachman. Decreased expression of caspase₃ in penis and prostate tissues of rat after treatment with buceng (*Pimpinella alpina* Molk) & *Eurycoma longifolia* Jack). Med J Indones. 2013;22:2-8.
- Godfrey B, Lin Y, Larson J, Haferkamp B, Xiang J. Proteasomal degradation unleashes the prodeath activity of androgen receptor. Cell Res. 2010;20(10):1138-47.
- 7. Frezza M, Yang H, Dou QP. Modulation of the tumor cell death pathway by androgen receptor in response to cytotoxic stimuli. J Cell Physiol. 2011;226(11):2731-9.
- 8. Youle RJ, Strasser A. The BCL-2 protein family: opposing activities that mediate cell death. Nat Rev Mol Cell Biol. 2008;9(1):47-59.
- 9. Bogner C, Leber B, Andrews DW. Apoptosis: embedded in membranes. Curr Opin Cell Biol. 2010;22(6):845-51.
- 10. Gump MJ, Thorburn A. Autophagy and apoptosis: what is the connection?. Trends Cell Biol. 2011;21(7):387-92.
- 11. Sharifi AM, Mottaghi S. Finasteride as a potential tool to improve mesenchymal stem cell transplantation for myocardial infarction. Med Hypotheses. 2012;78(4):465-7.
- 12. Libera DL, Ravara B, Gobbo V, Barbera MD, Angelini A, Vescovo G. Therapeutical treatments for blocking apoptosis and preventing skeletal muscle myopathy in heart failure. Basic Appl Myol. 2002;12(2):65-71.
- 13. Danial NN, Korsmeyer SJ. Cell death: critical control points. Cell. 2004;116(2):205-19.
- 14. Arnoult D. Mitochondrial fragmentation in Apoptosis. TRENDS in Cell Biology. 2006;17(1):6-12.
- 15. Valdivia I, Campodónico I, Tapia A, Capetillo M, Espinoza A, Lavín P. Effects of tibolone and continuous combined hormone therapy on mammographic breast density and breast histochemical markers in postmenopausal women. Fertil Steril. 2004;81(3):618-23.
- 16. Nieschlag E, Behre HM, Bouchard P, et al. Testosterone replacement therapy: current trends and future directions. Hum Reprod Update. 2004;10(5):409-19.
- 17. Barceloux DG, Palmer RB. Anabolic-androgenic steroids. Dis Mon. 2013;59(6):226-48.
- 18. Shittu LAJ, Shittu RK, Osinubi AA, Ashiru OA. Stereological evidences of epithelial hypoplasia of

seminiferous tubules induced by mesterolone in adult *Sprague-Dawley* rats. Afr J Endoc and Met. 2008;7:14-7.

- Gauthaman K, Adaikan PG, Prasad RN. Aphrodisiac properties of *Tribulus terrestris* extract (Protodioscin) in normal and castrated rats. Life Sci. 2002;71(12):1385-96.
- 20. Gauthaman K, Adaikan PG, Prasad RNV. Sexual effects of puncturevine (*Tribulus terrestris*) extract (protodioscin): an evaluation using a rat model. J Altern Complement Med. 2003;(9)2:257-65.
- 21. Meny S. Efek ekstrak purwoceng (*Pimpinella alpina* Molk) terhadap peningkatan biomarker fungsi ereksi pada tikus jantan *Sprague-Dawley* [dissertation]. Semarang: Diponegoro University; 2013. Indonesian.
- 22. Gauthaman K, Ganesan AP. The hormonal effects of Tribulus terrestris and its role in the management of male erectile dysfunction--an evaluation using primates, rabbit and rat. Phytomedicine. 2008;15(1-2):44-54.
- 23. Viktorov I, Bozadjieva E, Protich M, et al. Pharmacological, pharmacokinetic, toxicological and

clinical studies on protodioscin. IIMS Therapeutic Focus. 1994. Available from: http://www.libilov.com/ en/clinical_studies/study_IIMS_1994.htm

- 24. De Almeida FF, Andersson E, Mittelholzer C, Karlsen O, Taranger GL, Schulz RW. Pituitary gonadotropin and testicular gonadotropin receptor expression in Atlantic cod (*Gadusmorhua L*.) during the first reproductive season: Effects of photoperiod modulation. Gen Comp Endocrinol. 2011;173(1):111-9.
- 25. Kim HH, Schlgel PN. Endocrine manipulation in male infertility. Urol Clin North Am. 2008;35(2):303-18.
- 26. Manitto P. Biosynthesis of natural product. Connecticut. New Delhi: India; 1981. p. 236.
- 27. Morley JE, Malmstrom TK. Frailty, sarcopenia, and hormones. Endocrinol Metab Clin North Am. 2013;42(2):391-405.
- 28. Ohta Y, Nishikawa A, Fukazawa Y, Urushitani H, Matsuzawa A, Nishina Y, et al. Apoptosis in adult mouse testis induced by experimental cryptorchidism. Acta Anat (Basel). 1996;157(3):195-204.