Evaluation of PCR detection of S. typhi DNA in the diagnosis of clinically suspected Typhoid fever
A.B. Dey¹, Rama Chaudry², Maya Gopinath¹, D.S. Chandel², B.V. Laxmi²

INTRODUCTION
Enteric fever is a common public health concern in India and most developing societies¹,². This acute systemic disease is usually caused by infection with Salmonella typhi and to a lesser extent by Salmonella paratyphi. Typhoid fever is often marked by multi-systemic complications and death when diagnosed late or not treated effectively. With little possibility of its complete eradication, early diagnosis and effective treatment are the only means of reducing the impact of typhoid fever.

Isolation from peripheral blood by culture is the most definitive diagnostic procedure for detection of S. typhi infection. However the culture negativity rates are often as high as 30-65%³,⁴. Similarly, serological tests such as Widal test lacks diagnostic utility due to its non-specificity in endemic areas⁵,⁶. Attempts have been made in the past to develop better methods for diagnosis of the infection with limited success. Several procedures such as counter immuno-electrophoresis⁷, latex agglutination⁸ and coagglutination⁹ have been employed in the diagnosis of S. typhi infection with poor reproducibility. Application of ELISA techniques for detection of antigens of S. typhi and antibodies against it have also led to a major break through¹⁰,¹¹. In the last decade, of S. typhi by using DNA probes¹²,¹³ have been attempted in
clinical samples but has not lived up to their potential due to technical and cost factors. To improve the utility of detection of *S. typhi* DNA as a reliable tool, PCR amplifications has been applied in the diagnosis of typhoid fever in recent years with encouraging results\textsuperscript{15,17}. In the present communication, we have evaluated the application of PCR amplification of the dH flagellin gene in detection of *S. typhi* in patients with suspected enteric fever.

**MATERIALS AND METHODS**

**Cases**

In a prospective study, consecutive patients with acute febrile illness suspected to have enteric fever were investigated to achieve a definitive diagnosis during the period October 1995 and April 1997. The bases of a clinical diagnosis of enteric fever were i) continuous fever for at least 1 week duration, ii) gastrointestinal symptoms, iii) splenomegaly and/or hepatomegaly and iv) response to empirical therapy with ciprofloxacin or second or third generation cephalosporins within 72-96 hours. Patients were investigated according to a pre-designed protocol which included clinical evaluation (history taking and physical examination), routine laboratory investigations comprising of complete blood count, peripheral smear examination for parasites, liver function tests, routine blood chemistry, urinalysis and chest x-ray. Microbiological investigations included aerobic and anaerobic blood culture, urine culture, paired Widal test and PCR amplification and detection of *Salmonella typhi* DNA from flagellin gene.

**Table 1. Clinical features and laboratory investigations in sixty patients with suspected enteric fever.**

<table>
<thead>
<tr>
<th>Symptoms</th>
<th>Clinical features</th>
<th>No (%)</th>
<th>Laboratory investigations</th>
<th>No (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fever</td>
<td>60 (100)</td>
<td></td>
<td>Anemia (&lt;12 gm/dl)</td>
<td>15 (25)</td>
</tr>
<tr>
<td>Abdominal pain</td>
<td>12 (20)</td>
<td></td>
<td>Leucopenia (&lt;4000/mm(^3))</td>
<td>28 (46.7)</td>
</tr>
<tr>
<td>Diarrhoea</td>
<td>20 (33)</td>
<td></td>
<td>Leucocytosis (&gt;11000/mm(^3))</td>
<td>3 (5)</td>
</tr>
<tr>
<td>Constipation</td>
<td>5 (8.3)</td>
<td></td>
<td>Hyperbilirubinaemia (1.8 mg %)</td>
<td>1 (1.66)</td>
</tr>
<tr>
<td>Headache</td>
<td>38 (63.3)</td>
<td></td>
<td>Raised ALT (&gt;40U/dl)</td>
<td>52 (86.7)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Rectal bleeding</td>
<td>1 (1.7)</td>
<td></td>
<td>20 (33)</td>
</tr>
<tr>
<td>Vomiting</td>
<td>10 (16.7)</td>
<td></td>
<td>Raised AST (&gt;40U/dl)</td>
<td>53 (88.3)</td>
</tr>
<tr>
<td>Signs</td>
<td></td>
<td></td>
<td></td>
<td>26 (43.3)</td>
</tr>
<tr>
<td>Bradycardia</td>
<td>43 (71.7)</td>
<td></td>
<td>Albuminuria (2++)</td>
<td>10 (16.7)</td>
</tr>
<tr>
<td>Splenomegaly</td>
<td>54 (90)</td>
<td></td>
<td>Urine sediments (RBC, WBC)</td>
<td>12 (20)</td>
</tr>
<tr>
<td>Hepatomegaly</td>
<td>21 (35)</td>
<td></td>
<td>Abnormal chest x-ray</td>
<td>1 (3.3)</td>
</tr>
<tr>
<td>Abdominal tenderness</td>
<td>19 (31.7)</td>
<td></td>
<td>(miliary mottling)</td>
<td></td>
</tr>
<tr>
<td>Chest signs</td>
<td>11 (18.3)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lymphadenopathy</td>
<td>2 (3.3)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**PCR for *S. typhi***

Three millilitres of venous blood was collected in sterile citrate tube. DNA was extracted from 200 microlitre of blood using the Qiagen blood and body fluid protocol. The extracted DNA was subjected to PCR in the DNA thermocycler (MJ Research Inc Massachusetts, USA). The reaction mixture for PCR contained the extracted DNA, 50 picomoles each of RK1 (5'TGG GCG ACG ATT TCT ATG CC 3') and RK2 (5'TTT CGC GAA CCT GGT TAG CC 3'), 200 micromoles of all four dNTPs, 0.625 unit of Taq polymerase, 2.5 mM MgCl\(_2\) and standard PCR buffer to make a final volume of 25 microlitres. Denaturation was achieved at 94°C for 1 minute, annealing at 57°C for 1 minute 15 seconds and polymerisation at 72°C for 3 minutes. The PCR products were subjected to agarose gel electrophoresis, stained with ethidium bromide and examined under ultraviolet illumination. The specificity of RK1 and RK2 primers for dH flagellin gene has been reported in an earlier publication\textsuperscript{17}.

**Controls**

Blood samples from 21 afebrile controls were also subjected to Widal test and PCR amplification for *S. typhi* DNA.

**Analysis of data**

A diagnosis of *S. typhi* infection was considered when a case of suspected enteric fever demonstrated.
1. positive blood culture for *S. typhi*
2. a titre of ≥ 1:160 for O and H agglutinins on first test or four fold rise in O and H agglutinin titres on paired samples
3. demonstration of *S. typhi* DNA after PCR amplification.

RESULTS

Clinical manifestation

Sixty patients (40 males and 20 females, age range: 9 and 63 years) with acute onset of fever, suspected clinically to have enteric fever were investigated for a definitive diagnosis. The duration of fever at the time of presentation was 8.4 (± 2.3) days (range 7-18 days). Thirty seven patients had received various antimicrobial drugs which included cotrimoxazole, amoxycillin and ciprofloxacin for periods varying between 1 and 4 days. The clinical and laboratory manifestations of these patients are presented in Table 1.

Microbiological profile

With a diagnostic titre of 1:160 for both O and H agglutinins 11 patients had a positive Widal test at the testing. A repeat Widal test was carried out in 37 patients out of 45 cases (excluding 3 cases of typhoid, 5 cases of paratyphoid and 7 cases with non-enteric illnesses) in whom a diagnosis of enteric fever was considered after one week while getting treated for the same. Only two cases showed a four fold or more rise in titres of O and H agglutinins over their previous titres fulfilling the criteria for diagnosis of typhoid fever. Thus 13 cases in all had a positive Widal test. Results of Widal test in cases and controls are presented in Table 2.

In 13 cases, a putative causal organism was isolated from blood culture. They included *S. paratyphi* A in five, *S. typhi* in three, *Staphylococcus aureus* in two, *Escherichia coli* in one, *Klebsiella* species in one and *Enterococcus* in one. None of these cases had received pre-investigation antibiotic therapy. *Plasmodium vivax* was detected in peripheral blood smear in one patient. All the three cases who had *S. typhi* grown in their peripheral blood, had Widal test positive on first testing. Cases of paratyphoid had positive Widal test (high O agglutinin and high AH titres) where as all the five cases with septicaemia had non-diagnostic rise in O and H agglutinins.

Detection of *S. typhi* DNA by PCR amplification

*S. typhi* DNA could be detected in blood samples of seven cases. None of the blood samples from controls were positive for *S. typhi* DNA. One of them had blood culture positive for *S. typhi* and none of them had a positive Widal test on first testing but two of them had four fold or more rise in O and H agglutinins on paired serology. All the cases with septicemic illness other than typhoid were PCR negative.

The diagnosis arrived after the investigations are presented in Table 3. It was observed that nine of the...
seventeen typhoid cases had received prior antibiotic therapy. Of them five were diagnosed by PCR assay. None of the culture positive cases and four out of eleven Widal positive cases had been treated prior to hospitalisation.

**Treatment**

Forty six patients received ciprofloxacin, eight patients received cefotaxime and four patients were treated with cefuroxime axetil. Patients with malaria and miliary tuberculosis were provided specific treatment. All the cases responded to empirical/ specific antibiotic therapy and became afebrile between three and seven days (mean 5.2 ± 2.1 days). None of the patients had any complication of typhoid or reported relapse.

**DISCUSSION**

In a prospective study of sixty cases with acute onset fever suspected to have enteric fever, a diagnosis of typhoid was considered in 17 patients and paratyphoid in 5 patients. Other septicemic illness were detected in 5 patients. Malaria and tuberculosis was diagnosed in one patient each. In thirty one patients no definitive diagnosis (with microbiological evidence) could be achieved. Failure to reach a diagnosis in such large number of cases may be due to three factor i) cause of fever other than enteric fever i.e. other infections such as viral infections ii) effect of empirical antibiotic therapy prior to investigations and iii) lack of sensitivity of diagnostic procedures to detect *S. typhi* infection. Acute onset febrile illnesses in the community are usually infective in nature and their epidemiology is largely influenced by prevalence of various infectious diseases in a given geographical area. Causes of such fever include septicaemic infections with bacteria, known and unknown viruses, rickettsia and protozoa, and organs specific bacterial and viral infections of gastrointestinal tract, respiratory tract and urinary tract. The clinical manifestations of most short duration fevers in the initial phases are not distinctive as was observed in the present study. Empirical treatment of such illnesses with broad spectrum antibiotics by self or community physicians is not unusual often leading to complications, treatment failure and development of antibiotics resistance. The major reason for such empirical treatment is lack of quick and sensitive diagnostic procedures of the above mentioned infections. Diagnostic procedures in the context of enteric fever have been largely unsatisfactory due to low specificity of serological methods in endemic areas and low probability of isolation of *S. typhi* after antibiotic therapy. Newer techniques such as ELISA or DNA probes have not shown much clinical utility.

In the present study thirty one out of sixty suspected enteric fever cases remained undiagnosed from microbiology point of view and seven patients had a diagnosis other than typhoid or paratyphoid. Sixty percent of them had received some antimicrobial agent prior which probably affected the investigations in a large way. Of the 17 cases who were labelled as typhoid, nine had received antibiotics. As a result, culture positivity was extremely low. Widal test in the early phase of the illness could pick up eight cases. However, interpretation of the test remains difficult and the disease can be there even when the test is negative. Pre-investigation antibiotic treatment has probably a role as well in affecting the Widal test results. It was also observed that only two of the Widal negative patients had diagnostic titre in repeat testing a week later while receiving treatment where as in the large majority there was no such conversion. This reflects great individual variations in mounting an immunological response to the infection. PCR was positive in seven patient, of whom blood culture was positive in only one case. False negative PCR in two culture positive cases is difficult to explain and in earlier report we did not encounter this problem. The false negativity in these two cases is possibly due to denaturation of the sample due to problems of storage. Alternatively presence of mutant strains in blood sample can also lead to false negative result. In our earlier study we had suggested a specificity of 93.7% and sensitivity of 100% for PCR detection of *S. typhi* in typhoid. However, when applied in clinical situation, these figures have not been substantiated. Larger studies are required in testing the value of the test especially in presence of prior antibiotics therapy. In the present study five of the seven PCR positive cases had prior antibiotic therapy and all of these were culture and Widal negative in the first testing. Thus PCR detection of *S. typhi* is a useful tool in the diagnosis of enteric fever in situations of prior antibiotic therapy. As PCR assay also requires a critical quantity of DNA in the blood sample, PCR positivity is also likely to be affected by the duration of antibiotic therapy which results in rapid decline in the load of susceptible infectious agent.

**CONCLUSION**

Definitive diagnosis of typhoid is difficult to achieve especially in antibiotic treated patients. PCR detection of *S. typhi* appears to be a useful tool in the di-
agnosis of typhoid fever in patients with prior anti-
biotic therapy.

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