

Poor treatment compliance leads to a higher mutation for rifampicin resistance in multibacillary leprosy patients

Yulia Siskawati, Evita H. Effendi, Lili Legiawati, Sri L. Menaldi

Department of Dermatovenereology, Faculty of Medicine, Universitas Indonesia, Cipto Mangunkusumo Hospital, Jakarta, Indonesia

ABSTRACT

Background: Multidrug therapy (MDT) is a safe and effective drug combination for leprosy treatment that can prevent drug resistance. *Mycobacterium leprae* resistance, especially to rifampicin, is a serious problem as it potentially thwarts the worldwide leprosy-elimination program by the World Health Organization (WHO). One of the suspected causes of rifampicin resistance is poor treatment compliance. It was necessary to assess the association between the treatment compliance and the occurrence of mutation rifampicin resistance in multibacillary (MB) leprosy patients.

Methods: A comparative, analytical, cross-sectional study was performed in MB leprosy patients who had completed treatment at the Dermatovenereology Outpatient Clinic in Cipto Mangunkusumo Hospital and the Sitanala Center for Leprosy Hospital from October 2012 to April 2013. Based on treatment regularity and history of drug discontinuation, the subjects were classified as either having good or poor compliance. Skin smear from a slit skin smear (SSS) examination was further analyzed by using the polymerase chain reaction (PCR) sequencing technique to detect rifampicin resistance.

Results: Fifty-seven study subjects were enrolled in this study. In the good treatment compliance group (29 subjects), only 1 case of mutation for rifampicin resistance was found. Meanwhile, in the poor drug compliance group (28 subjects), 8 cases of mutation for resistance (29%) were found. This difference in mutation rate was statistically significant (OR=11.2; 95% CI=1.296–96.787; p=0.012).

Conclusion: This study revealed that the risk of occurrence of *M. leprae* resistance to rifampicin in patients with poor drug compliance was significantly higher than in those with good drug compliance.

Keywords: multibacillary leprosy, rifampicin resistance, treatment compliance

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Corresponding author: **Yulia Siskawati**
yuliasiska@gmail.com

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Leprosy is a chronic infectious disease caused by *Mycobacterium leprae* (*M. leprae*), which initially affects peripheral nerves and subsequently extends to nearby skin tissue, the oral mucosa, upper respiratory tract, reticuloendothelial system, eyes, muscles, bones, and testicles.¹ Previously, dapsone was the only antibiotic given to treat leprosy. However, emerging resistance to this monotherapy strategy has led to the introduction of multidrug therapy (MDT) by the World Health Organization (WHO) in 1981.²⁻⁴ This MDT is a combination of drugs proven to be safe and effective to treat leprosy.⁵ For multibacillary (MB) leprosy, MDT consists of rifampicin, dapsone, and clofazimine, whereas for paucibacillary (PB) leprosy, it consists of rifampicin and dapsone.⁶ It is well known that administering 2 or more antibiotics with different mechanisms of action in combination can help to prevent drug resistance.³ Resistance to MDT, especially rifampicin, warrants special attention because it is a potential cause of failure of the worldwide leprosy-elimination program by the WHO.⁷

Rifampicin is a strong bactericidal antibiotic effective against both gram-positive and gram-negative bacteria.⁸ Single-dose of rifampicin is proven effective at eradicating 99.99% of *M. leprae*.⁹ Its bactericidal effectivity works through inhibition of DNA-dependent RNA polymerase, which is coded by the *rpoB* gene. Therefore, any mutation within the gene and/or its associated gene(s) could result in a conformational change in the polymerase that hinders rifampicin adherence.¹⁰ Another possible mechanism of rifampicin resistance is a change in bacteria cell wall permeability, which reduces the total amount of rifampicin available in the cell.^{8,10}

Resistance to any component of leprosy MDT is confirmed by *M. leprae* inoculation on the sole of a guinea pig foot or by biomolecular study using polymerase chain reaction (PCR).¹¹ This biomolecular study can detect specific drug-resistance-causing gene mutations.¹² The mutation can be detected even if the specimen contains a low bacterial load.¹³

A study conducted by Maeda et al¹⁴ in 2001 encompassing several countries such as Japan, Haiti, Indonesia, Pakistan, and Philippines showed that 13 of 88 isolates (14.8%) are mul-

tidrug-resistant *M. leprae*. In 2007, Matsuoka¹¹ found that from 252 isolates, rifampicin resistance in relapse cases reached 20% in Maluku Utara, Indonesia and Sulawesi Utara, Indonesia, while it was only 8.3% in Yangon, Myanmar. In this study, isolates were taken from all leprosy patients who fulfilled the inclusion criteria, regardless of completion of therapy.⁷

There are two possible etiologies for rifampicin resistance. Firstly, resistance is correlated with a previous history of rifampicin monotherapy.^{8,15} Secondly, it is related to poor compliance, including self-regulated drug administration and early treatment withdrawal.¹⁵ Several reasons underlying poor compliance are physical limitations (elderly and disability/body deformity), inability to visit the physician during working hours, drug side effects, doubt in drug efficacy, social stigma, lack of reliable transportation, and location of the health care center far from home. However, Kar et al¹⁶ conducted a retrospective cohort study related to MDT compliance in 254 leprosy cases from 2002 to 2005 in India, and this study found that the poor compliance rate was higher in large urban centers, even though health care centers are readily available and the patients are much better educated than in rural areas.

This study was aimed to evaluate the association between rifampicin resistance and poor treatment compliance in MB leprosy patients after completion of treatment.

METHODS

This is an analytic, comparative, cross-sectional study in MB leprosy patients who had completed a treatment course.

Subjects and sample size

Multibacillary leprosy patients at the Dermatovenereology Outpatient Clinic of the Cipto Mangunkusumo Hospital (PKK-RSCM) and Sitanala Center for Leprosy Hospital Tangerang with a positive slit skin smear (SSS) bacterial index who had completed an MDT treatment course were included in the study. Based on the previous study, the rifampicin-resistant mutation prevalence was 44% in the poor treatment compliance group and 14% in the good treatment compliance group.¹⁵

Table 1. Baseline characteristics of subjects

Characteristics	Compliance		Total n (%)
	Good (n=29) n (%)	Poor (n=28) n (%)	
Gender			
Male	21 (72.4)	24 (85.7)	45 (78.9)
Female	8 (27.6)	4 (14.3)	12 (21.1)
Age group (years)			
15–24	3 (10.3)	9 (32.2)	12 (21.1)
25–44	15 (51.7)	13 (46.4)	28 (49.1)
>44	11 (38.0)	6 (21.4)	17 (29.8)
Educational background			
Low	12 (41.4)	16 (57.1)	28 (49.1)
High	17 (58.6)	12 (42.9)	29 (50.9)

Therefore, the minimum number of subjects in each group using the sample size formula for proportion comparison was 28 subjects.

Study procedure

Patients who fulfilled the clinical eligibility criteria underwent SSS laboratory examination to determine if they met the positive bacterial index criteria. Three locations for SSS specimen collection were chosen according to previous SSS examination during the early evaluation. The blade which had been utilized to take a smear from each location was placed into an Eppendorf® tube for the *M. leprae* DNA suspension and extraction process using the QIAGEN QIAprep® Spin Miniprep Kit. Each specimen containing extracted *M. leprae* DNA underwent the next step, which was *rpoB* gene amplification, initially by PCR. Specimens showing negative results underwent subsequent nested PCR. Both PCR processes were carried out using the Takara® PCR Thermal Cycler (model TP 600). Finally, the amplified target gene was sequenced by using Dual CyCye™ Terminator Sequencing Kits (ABI) to detect the specific nucleic acid arrangement.

Evaluation technique

Mutations in the *rpoB* gene that could cause significant resistance to rifampicin were defined as any type of mutation found in gene codon numbers 407, 416, 420, 425, and 427; or one amino acid insertion between codon numbers 408 and 409.¹⁷ At the same time, the subjects

were classified into 2 groups based on the degree of treatment compliance, either good or poor. The group with poor treatment compliance included all subjects with a history of drug withdrawal for a period at certain point(s) during the MDT treatment course but in the end completed it, or irregular dapsone and clofazimine consumption defined as at least 7 days of not consuming both drugs in the same month.

Ethical considerations

This study passed ethical evaluation according to the standards of the Ethics Committee of the Faculty of Medicine, Universitas Indonesia and Research Division of Cipto Mangunkusumo Hospital (603/H2.F1/ETIK/2012). These standards were in accordance with those of the Helsinki declaration. All patients were agreed and gave informed consent for this study.

Data processing and analysis

All data were recorded and coded to be processed statistically using Statistical Product and Service Solutions (SPSS) 20.0 for Windows. The resistance rate was calculated by Chi-square test and presented as an odds ratio (OR) and 95% confidence interval (CI).

RESULTS

Subject characteristics

During a 7-month sampling period from October 2012 to April 2013, 57 patients were eligible for the study, 41 of which were from PKK-RSCM and the rest were from Sitanala Center for Leprosy Hospital. Based on their compliance with treatment, there were 29 subjects with good compliance and 28 subjects with poor compliance. The baseline characteristics of these patients following classification into these groups is depicted in Table 1.

Several treatment history characteristics were evaluated. Regarding compliance, 97% of subjects with good compliance and 32% of those with poor compliance visited a healthcare center of at least once monthly. Specifically, monthly rifampicin consumption was adhered to in 100% and 79% of cases in the good and poor compliance groups, respectively. There was no history of rifampicin monotherapy in any subject from either group.

Clinically, 74% of all subjects were categorized as the "Borderline" (BL) type based on the Ridley & Jopling classification scheme. The distribution of each leprosy type by this classification between good and poor treatment compliance was almost identical.

PCR result and mutation pattern of the *rpoB* gene

Electrophoresis of the *rpoB* gene PCR product is shown in Figure 1. Of the 57 subjects, 48 were positive by PCR (PCR positivity rate 84%). All specimens with positive PCR results were subjected to the sequencing process to detect point mutations related to rifampicin resistance. Point mutations were detected in 9 samples (16%). The most common mutation found in 8 samples was at codon number 410, which encodes aspartic acid (GAT) in wild-type *M. leprae* but encodes tyrosine (TAT) in rifampicin-resistant mutant *M. leprae*, followed by mutation at codon number 425 (TCG encoding serine mutated into TTG encoding leucine) found in 1 sample (Figure 2). Additionally, there were 3 specimens with silent mutations: in 1 case, codon 420 CAC was mutated to CAT, both of which encode histidine, while in 2 other cases, codon 412 AAC was mutated to AAT, both of which encode asparagine.

Comparison of rifampicin resistance between patients with good and poor treatment compliance

Out of 29 subjects with good treatment compliance, there were 25 samples (86%) positive by PCR, while positive PCR results were found in 23 (82%) out of 28 subjects with poor treatment compliance. Subjects with negative results were still included in the analysis and considered as not showing rifampicin resistance. When a sample is negative by PCR, it is assumed that there is no bacterial DNA detected in the sample, and no rifampicin resistance will be found. A statistical comparison of rifampicin resistance prevalence between the 2 groups is shown in Table 2.

DISCUSSION

Clinical characteristics of subjects

The ratio of male-to-female subjects was 3.75:1. These data were in agreement with a global epidemiology study which found a male bias in leprosy patients with the ratio of males to

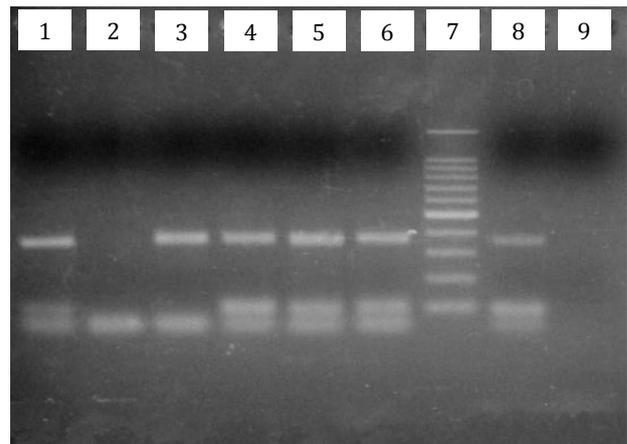


Figure 1. Electrophoresis result of *M. leprae rpoB* gene PCR from a slit skin smear (SSS) specimen. Wells number 1, 3, 4, 5, and 6, samples positive by PCR. Well number 7 was a 100-bp DNA ladder. Well number 8 was a positive control, and number 9 was a negative control

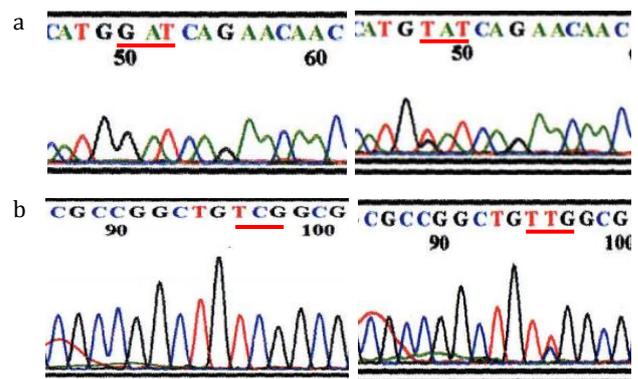


Figure 2. Nucleotide arrangement as a result of *rpoB* gene sequencing in: (a) codon number 410 showed a wild-type (GAT, left) and mutation (TAT, right); (b) codon number 425 showed a wild-type (TCG, left) and mutation (TTG, right)

females ranging from 1.5 to 2:1.^{18,19} In addition, according to leprosy morbidity data at PKK-RSCM in 2012, there were 152 (70.4%) and 64 (29.6%) new male and female leprosy patients, respectively. The male-to-female ratio reached 2.4:1.²⁰ The majority (49.1%) of subjects were in the 25- to 44-year age group, the distribution of which was bimodal, one mode is the 35- to 44-year-old group.²¹ The previous PKK-RSCM leprosy data registry in 2012 showed an almost identical result: 49.2% of MB leprosy cases were in the 24- to 44 year-old age group.²⁰ Lastly, in this study, nearly half of the subjects (49.1%) had a low educational level. After classification based on the degree of compliance, the difference in 1) male-to-female ratio, 2) age distribution, and 3) educational level between poor and

Table 2. Comparison of rifampicin resistance based on treatment compliance in MB leprosy patients

Treatment compliance	Mutation for resistant n (%)	Non-resistant n (%)	p	OR (95% CI)
Poor	8 (89)	20 (42)		
Good	1 (11)	28 (58)	0.012	11.2 (1.296–96.787)
Total	9 (100)	48 (100)		

MB=multibacillary; OR=odds ratio; CI=confidence interval

good treatment compliance were not sufficient in magnitude to cause any significant bias in interpreting the analytical part of the study.

The MDT regimen for MB leprosy consisted of monthly (every 28 days) rifampicin and clofazimine (300 mg) that should be administered daily and under the supervision of a physician, self-administered dapsone and clofazimine (50 mg). Accordingly, the lower frequency of monthly visits to a healthcare center in those with poor compliance predicted lower monthly rifampicin consumption, one of the significant differences found in this study regarding treatment history characteristics. These data showed that the pre-determined criteria for the poor compliance group are aligned with rifampicin consumption regularity, which is presumably related to its resistance. Another major risk factor for rifampicin resistance is a history of rifampicin monotherapy, which was not found in any of the subjects included in this study. Therefore, history of rifampicin monotherapy could be excluded as the cause of resistance if any resistant isolate were to be found.

The dominance of the BL leprosy type in this study was commensurate with the PKK-RSCM leprosy data registry in 2012, which found the BL type to be the most common (50.9%) among all new leprosy cases.²⁰ Other than clinical manifestations, the leprosy classification by Ridley & Jopling also reflects the bacterial index.¹ Supposedly, the almost identical distribution of leprosy types between good and poor treatment compliance would be followed by similar bacterial indices. Nevertheless, this study found a significant difference in bacterial index distribution between the 2 groups. The reason behind this finding was beyond the scope of this study and

warrants further research. Fortunately, it did not appear to affect the PCR positivity rate, as this rate was similar in both groups.

Mutation pattern of *rpoB* gene

Not all specimens were positive by PCR. This finding could have resulted from fragmented *M. leprae* DNA that rendered it impossible to be detected through PCR, in which the primer used for *rpoB* gene consisting of 337–358 bp. Other possibilities were error in specimen handling during transportation from Jakarta to Surabaya and differences among laboratory workers' experience and expertise.

The prevalence of significant mutations related to rifampicin resistance in this study was quite high, reaching 16%. Wahyuni et al²² reported only 1 case of rifampicin resistance among 270 isolates which were collected from 2003 to 2011. This prominent difference possibly occurred because of the difference in inclusion criteria; in this study, all subjects had completed the MDT course and had positive bacterial index results.

There were only 2 types of point mutation in this study: the most common being mutation of codon 410 GAT (aspartic acid) to TAT (tyrosine), followed by mutation of codon 425 TCG (serine) to TTG (leucine). A previous study by Matsuoka et al¹⁵ found that the most common mutation was codon 425 TCG (serine) to TTG (leucine), with a total number of 6 cases, followed by mutation in codon 410 GAT (aspartic acid) to TAT (tyrosine), codon 420 CAC (histidine) to GAC (aspartic acid), and codon 425 TCG (serine) to ATG (methionine), with 1 case each. Meanwhile, the only rifampicin-resistant case found in the study conducted by Wahyuni et al²² resulted from mutation of codon 410 GAT (aspartic acid) to TAT (tyrosine).

To date, there has not been a single report about the causal association between the point mutation found in this study and rifampicin resistance. To prove definitively whether the *M. leprae* isolates in the specimens were resistant to rifampicin, the gold standard diagnostic procedure was the inoculation of *M. leprae* on the foot of a guinea pig and observe its response to rifampicin treatment. Clinically, these leprosy patients whose isolates showed a point mutation need re-evaluation to detect a possible concurrent rifampicin-resistance-related mutation. This suggestion was based on other studies in which the samples showed a point mutation in codon unrelated to rifampicin resistance. After re-evaluation, it was found that there was actually concomitant mutation of a codon related to rifampicin resistance.^{23,24}

Association between treatment compliance and rifampicin resistance

There was a statistically significant relationship between treatment compliance and the occurrence of rifampicin resistance ($p=0.012$). MB leprosy patients whose treatment compliance was poor had a significantly higher risk of developing resistance to rifampicin than those whose compliance was good. A previous study by Matsuoka et al¹⁵ in 2007 reported that rifampicin resistance prevalence was as high as 20% in relapsing patients. One of the risk factors of leprosy relapse was poor compliance in previous treatment, and this study shows that high rifampicin resistance was related to treatment compliance. The rifampicin resistance rate in patients with poor compliance reached 89%, while it was only 11% in those with good treatment compliance.

In conclusion, poor treatment compliance was associated with a higher risk of rifampicin resistance in MB leprosy patients.

Conflicts of interest

The authors affirm no conflict of interest in this study.

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