Genetic polymorphism of merozoite surface protein-1 (MSP-1) block 2 allelic types in *Plasmodium falciparum* field isolates from mountain and coastal area in West Sumatera, Indonesia

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**Abstract**

**Background:** The field isolates of *P. falciparum* may display variant forms and different frequencies. This study was designed to know the diversity of allelic type of MSP-1 block 2 among *P. falciparum* isolates collected in a mountain and a coastal area in West Sumatera, Indonesia, and compare mountain and coastal area.

**Methods:** A total of 56 *P. falciparum* infected blood samples, collected from 27 patients attending local health facilities in South Solok district in a mountain region and 29 patients attending a local health facilities in South Coastal district region, West Sumatera, Indonesia were used in this study. The regions flanking the highly polymorphic characters, block 2 for MSP-1, were genotyped by allele-specific nested-PCR to analyse the population diversity of parasite. Sequence analysis of the polymorphic regions of MSP-1 was also conducted to identify allelic diversity in the parasite population.

**Results:** Diverse allelic polymorphism of MSP-1 was identified in *P. falciparum* isolates from a mountain area and a coastal area in West Sumatera, Indonesia, and most of the infections were determined to be mixed infections. Sequence analysis of MSP-1 block 2 revealed that 16 different alleles for MSP-1 (3 for K1 type, 2 for MAD20 type and 2 for RO33 type) were identified.

**Conclusion:** Extensive genetic polymorphism with diverse allele type was identified in MSP-1 in *P. falciparum* field isolates from a mountain and a coastal area. A high level of mixed infections was also observed, as was a high degree of multiplicity of infection. (Med J Indones 2011; 20:11-4)

**Key words:** allelic types, coastal area, mountain area, MSP-1 block 2, *Plasmodium falciparum*

Genetic diversity displayed by *P. falciparum* field isolates, the occurrence of variant forms of parasite at different frequencies in different geographic areas, and the complexity of infection represent major obstacles for the effective control of malaria. The propagation of multi-drug resistant parasites and insecticide-resistant mosquitoes has led to major difficulties in controlling the spread of malaria. To fight against malaria, an effective vaccine is urgently needed. A number of antigens expressed at different stages of parasite’s life cycle have been characterized with respect to their use in vaccine development against *P. falciparum*. Merozoite surface protein-1 (MSP-1) is one of the most promising vaccine candidates. People naturally exposed to *P. falciparum* develop antibodies against MSP-1. Furthermore, an association between a naturally acquired immune response to MSP-1 and reduced malaria morbidity has been observed. In a number of independent studies,
immunization with purified native MSP-1 or a recombinant fragment of protein has induced at least partial protection against parasite challenge. Sequence comparisons showed that the entire MSP-1 gene could be divided into 17 blocks that are variable, conserved, or semiconserved.4 In seven blocks, 1, 3, 5, 12, and 17, the sequences are conserved. In seven blocks (blocks 2, 4, 6, 8, 10, 14, and 16), the sequence show extensive diversity, while in the remaining (blocks 7, 9, 11, 13, and 15), the sequence are semiconserved. Variation in sequences of variable regions are dimorphic (K1/Wellcome or MAD20) in nature with the exception of the trimorphic–encoding region in block-2, which has a third version (RO33) found in natural isolates.5,6 Naturally acquired antibodies react more frequently against variable rather than conserved MSP-1 blocks and are specific for one of the major versions of variable blocks.7

In the current study, we have analyzed polymersase chain reaction (PCR)-amplified fragments containing variable blocks 2, 4, and 12-16 of the MSP-1 gene in the P. falciparum natural population and allelic types were scored by sampling allele-specific radio-labeled oligonuclotide probes. The allelic types were compared among the isolates collected from regions of hyperendemic malaria transmission (RHEMT) and mesoendemic malaria transmission (RMEMT).8,9 We have also analyzed the allelic diversity in the isolation showing more than strain of parasites.

METHODS

Clinical samples

The study was conducted in South Solok district, a area located in Bukit Barisan Mountain and in Pesisir Selatan, a area located in west coastal in West Sumatera province, Indonesia. The area has a farming of cocoa and sawit coconut in South Solok and majority was the fishing community in Pesisir selatan. All area is infested a primary vector, Anopheles balabacensis in South Solok and Anopheles sundaeicus in Pesisir Selatan and is characterized by high altitude, relative high humidity, constant rain, and an average of temperature 23°C, in South Solok, and coastal area, relative high humidity, constant rain, and an average of temperature 30°C in Pesisir Selatan. The analyzed samples were collected during an outbreak in which the prevalence of malaria was 60%, measured as the percentage of people with parasites among those who presented with malaria symptoms at a health service.

The samples was collected by finger puncture in the form of thick smears on slides. The slides were stained with Giemsa, and the presence of P. falciparum was detected under microscopic observation. The slides were then sent to the Medical Faculty, Andalas University in Padang, and parasitemia was determined and normalized for 100 leukocytes. This information was converted into the number of parasites per microliter, assuming a leucocyte count of 8,000/μL. To prepare DNA of the clinical samples, the thick smear, moistened previously with 1% saponin, the incubated for one hour at 4°C. The sample was centrifuged at 12,000 xg for five minutes, and the supernatant discarded. The precipitate was resuspended in 40 μL of Chelex-100 5% and boiled for 10 minutes. Finally the sample was centrifuged at 12,000 xg for five minutes, and the supernatant was recovered and stored at 4°C.10,11 We used 8 μL of the extract to amplify the MSP-1 gene through polymerase cian reaction (PCR).

The first amplification were used as a template for respective nested PCR assays. We took 5 μL to identify the allotypes in block 2 of MSP-1.

Allotype detection in block 2 of MSP-1

In the PCR-based amplification for MSP1 gene and the nested PCR to identify the allotypes in block 2 of this gene, we used an amplification profile with an initial denaturation at 94°C/2 min followed by 72°C/2 min, time during which the Taq DNA Polymerase enzyme was added. Then, 35 cycles at 94°C/30 sec, 55°C/30 sec, and 72°C/2 min were performed, ending with a final extension at 72°C/2 min. The following primers were used in amplifying the MSP1 gene: OK11 (5′ TAG AAG ATG CAG CAG TAT TGA CAG GTT A 3′) and OK12 (5′ ATT CTA ATT CAA GTG GTG CAT CAG TAA ATA A 3′). To define the allele present in block 2 of the MSP-1 gene, the primers OK1 (CTT AAA TGA TGG AGC AAG AAAT TAC TAC TAC AAA AGG TGC 3′) and OK2 (5′ GAG GGC TTG CAC CAG ATG AAG T 3′) were used for the K1 allelic type. The primers OK3 (5′ GTA TTA GAA GGA ACA AGT GGA ACA A 3′) and OK4 (5′ TAT CTA AAG GAT TGG TAT TAC GTC TGT AAC TAT T 3′) were used to typify the MAD20 allotype. The primers Primer-primer OK5 (5′ ATT AAA GGA TGG AGC AAA TAC TCA TGT TGT T 3′) and OK6 (5′ TCA TGC GGA TTT GCA GCA CCT GGA GA A 3′) were used to amplify the RO33 allelic type. Both the primary and the nested PCR were conducted in a final volume of 50 μL, using 200 μM of each dNTP, 1 μM of each primer, 2.5 U of TaqDNA polymerase (Promega) per reaction and 1 mM of MgCl2 in the enzyme buffer (50 mM KCI, 10 mM Tris-HCl, pH 9.0, 0.1% Triton x100).
Allelic distribution and complexity of infection

The prevalence of each allelic type analyzed was determined as the percentage of PCR fragments for the type in the total number of amplified bands for the corresponding locus. The complexity of infection, which is the average number of PCR bands per infected individual, was determined as described earlier. The percentage for type and complexity of infection were calculated independently for each genetic marker.

RESULTS

Of 56 samples analyzed from mountain area and coastal area, the endemic area in West Sumatera result in MSP1 was 14 peoples respectively. Genetic diversity was analyzed through PCR amplification of polymorphic regions in MSP1 gene. The three previously reported allelic types were found in block 2 of MSP1 from 28 samples were K1, MAD20, and RO33 type with the result as following: (Table 1)

<table>
<thead>
<tr>
<th>Combined allelic types</th>
<th>Mountain area</th>
<th>Coastal area</th>
</tr>
</thead>
<tbody>
<tr>
<td>K1</td>
<td>0</td>
<td>2</td>
</tr>
<tr>
<td>MAD20</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>RO33</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>K1,MAD20</td>
<td>3</td>
<td>7</td>
</tr>
<tr>
<td>K1,RO33</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>MAD20,RO33</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>K1, MAD20, RO33</td>
<td>10</td>
<td>4</td>
</tr>
<tr>
<td>Total</td>
<td>14</td>
<td>14</td>
</tr>
</tbody>
</table>

These types were established on the basis of differences in size observed through electrophoresis and showed respective sizes of 99-147 bp, 110-136 bp, and 99-120 bp of K1, MAD20, and RO33 respectively. The allelic most often present in the parasite population of the samples in mountain area studied was MAD20, with the frequency of 36.84% (14 of 38), K1 34.21% (13 of 38), and RO33 28.94% (11 of 38). Similarly, the allelic most often present in the parasite population of the samples in coastal area studied was K1, with the frequency of 44.8% (13 of 29), MAD20 37.93% (11 of 29), and RO33 17.24% (5 of 29).

<table>
<thead>
<tr>
<th>Located</th>
<th>K1</th>
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<tbody>
<tr>
<td></td>
<td>99 bp</td>
</tr>
<tr>
<td>Mountain area</td>
<td>3</td>
</tr>
<tr>
<td>Coastal area</td>
<td>3</td>
</tr>
</tbody>
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DISCUSSION

The structure of natural P. falciparum population plays a highly important role in the natural acquisition of immunity in malarial infection. Knowledge of this structure is necessary to develop strategies to control the disease, beginning with the design of effective vaccines against P. falciparum and including policy on the use of antimalarial medicines. We analyzed the genetic diversity of P. falciparum isolates collected in mountain endemic area and in coastal endemic area in West Sumatera, Indonesia for malaria but one where there is no permanent transmission of the parasite, as in certain African zones.

From the result was known that there were three infection combinations in mountain area and four infection combinations in coastal area. This condition may be caused by both these study areas were the open areas for all the comers and result in chance to occur mutation to P. falciparum infection is very large that is genetic recombination and variable result. Although the mountain and the coastal area in West Sumatera is the low endemic area, in Sumatera allelic variation is highly. This is much the same to result obtained in other region, such as Thailand and Senegal. The presence of novel variation is due to transmigration from other island in Indonesia to Sumatera terrestrial is the important issue.

In this study is also found amount balance of K-type allele, MAD20-type allele, and RO33-type allele in their frequency. This condition is proven with other regions having geographical isolation. For example, the studies by Diana Gomez only find MAD20 and K-type alleles, by fragment variance is restricted. Because there are in isolation areas in Colombia.

In some same research in different geographic area than msp1 block 2 as a marker, report important
variation in the genotype frequency. For example, in the other studies were found similar MAD20, K1, and RO33 allelic frequencies. This is highly different than the studies in France Guiana15 that only find a little MAD20-type allele and K1–allele type is plenty.

In this study is also found the fragment length of each allele. This variation and condition demonstrate the diverse MSP1 block 2 gene in each region, such that it cannot as pattern to other regions. The geographic factor very determine diversity of MSP-1 gene.

When various genotypes to a locus–parasite specific is found in a host, the possibility cross–fertilization and meiotic recombination in vector mosquitoes rise. In some level, the genetic diversity is expected to depend on the proportion of mixed infection and amount of clone per individu. The presence of mixed infection is one of prerequisite if the cross-fertilization is to produce new generation and enhance to diversity of parasite population.

REFERENCES