Resistance patterns of microbes isolated from gastrointestinal tract

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Abstract

Background: Infection of the gastrointestinal tract is one of important medical problems worldwide, especially in children. In developing countries such as in Asia, Africa and Latin America diarrheal diseases in children cause 12,600 deaths/day. The highest morbidity rates were found in children within 6-11 month age group; the mortality rates were highest in infants under 1 year old and children of one year old. Based on the estimation of 1980 population, the yearly morbidity and mortality in children under 5 years old in Africa, Asia (excluding China) and Latin America were 744-1000 million episodes and 4.6 million respectively. Recent report stated that the incidence of diarrhea for children under 5 years old in developing areas and countries was a median of 3.2 episodes/child/year, whilst the estimation of mortality rate was 4.9 children/1000/year. Improvement and advances in diagnosis and treatment of diarrheal diseases was done continuously, though they still cannot be reached by many developing countries in the world.

Gastrointestinal infections can be caused by bacteria, virus, fungi and parasites. Specifically for bacterial infection of digestive system, there are two...
major mechanisms. First, intoxication, the toxin is produced, ingested and cause symptoms. One of the most common organisms that cause intoxication is enterotoxin producing *Staphylococcus aureus*. Second, actual infection of the host cells, the pathogens produce toxin and result in host cell damage or death. Some organisms only adhere to the surface of epithelial cells and cause damage or death of the cells e.g. Enterotoxigenic *Escherichia coli* (ETEC). Others, after adhering, invade the cells and then damage them e.g. Campylobacter, Salmonella, Shigella, Rotavirus, and Norwalk agent. Microbes can produce the symptoms of watery diarrhea (viral gastroenteritis) to bloody mucus (dysentery, Shigellosis), as well as invade into the bloodstream as enteric fever by *Salmonella typhi*. Therefore, to support the diagnosis and management of bacterial infection, we conducted a study to determine the types of bacteria that were isolated from the gastrointestinal tract and their antibiotic resistance patterns.

**METHODS**

The samples were collected from feces, rectal or anal swab of patients who came to the Clinical Microbiology Laboratory FKUI during 2005-2008. Rectal/anal swabs were mostly collected from healthy patients for the screening of *Salmonella typhi* carrier.

Isolation, identification and antibiotic sensitivity test were conducted according to the standard operational procedure applied in the laboratory.

**Isolation**

To isolate microorganisms, we streaked the specimens into enteric medium (eosin methylen blue and Salmonella-shigella agar). The medium was then incubated at 37°C, for 18-24 hours. Gram stain was only conducted to identify colonies that grow on the medium if necessary (mostly rod negative Gram).

**Identification**

To identify microbes into species level, we conducted conventional biochemical tests. Biochemical tests were based on carbohydrate fermentation, enzyme utilization and other chemical substance reaction/production. If there was any problem in identifying the isolated microbes, we conducted Microgen™ GN-ID test (a commercially available diagnostic kit) to identify the microbes.

**Antibiotic sensitivity tests**

Disc diffusion method, which is a method of choice for testing the sensitivity of some antibiotics against microorganisms, was used to test the microorganisms that were isolated from the specimens. We used some antibiotics, such as: amoxicillin, sulbenicillin, cefotiam, ceftriaxone, amikacin, chloramphenicol, trimethoprim/sulfamethoxazole, ciprofloxacin, ofloxacin, cefepime, fosfomycin, piperacillin/tazobactam, levofloxacin, cefoperazone/sulbactam, meropenem and ticarcillin. The interpretation was based on the diameter of inhibition zone, which was seen as a clear zone that showed the ability of the antibiotics to inhibit the growth of the microbes. Antibiotic sensitivity test results were interpreted using NCCLS/CLSI guidance.

**Data collection and analysis**

The data collected were the types of isolated bacteria and their resistance patterns against some antibiotics. All data were noted, tabulated and presented in a graph. The data was analyzed using WHOnet version 5.3., to compute the percentages of antibiotics’ resistance and presented in a graph.

**RESULTS**

The result showed that pathogenic *E. coli* was the predominant isolate found. Table 1 shows the microbes that were isolated from feces, rectal or anal swab. In 2007-2008, the numbers of samples were higher because we examined rectal swab/feces that were collected from hospital’s and hotel’s employees.

<table>
<thead>
<tr>
<th>Year</th>
<th>Organisms</th>
<th>Number of samples</th>
<th>Percentage (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2005</td>
<td>Pathogenic <em>E. coli</em></td>
<td>2</td>
<td>1.5</td>
</tr>
<tr>
<td></td>
<td><em>S. paratyphi A</em></td>
<td>1</td>
<td>0.8</td>
</tr>
<tr>
<td></td>
<td>No pathogenic bacteria</td>
<td>127</td>
<td>97.7</td>
</tr>
<tr>
<td>2006</td>
<td>Pathogenic <em>E. coli</em></td>
<td>8</td>
<td>7.8</td>
</tr>
<tr>
<td></td>
<td>No pathogenic bacteria</td>
<td>95</td>
<td>92.2</td>
</tr>
<tr>
<td>2007</td>
<td>Pathogenic <em>E. coli</em></td>
<td>6</td>
<td>3.7</td>
</tr>
<tr>
<td></td>
<td>Yeast</td>
<td>1</td>
<td>0.6</td>
</tr>
<tr>
<td></td>
<td>No pathogenic bacteria</td>
<td>156</td>
<td>95.7</td>
</tr>
<tr>
<td>2008</td>
<td>Pathogenic <em>E. coli</em></td>
<td>12</td>
<td>3.9</td>
</tr>
<tr>
<td></td>
<td>Yeast</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>No pathogenic bacteria</td>
<td>290</td>
<td>95.1</td>
</tr>
</tbody>
</table>

Figure 2 shows the resistance pattern of pathogenic *E.coli* against several antibiotics. From 2005-2008, the pathogenic *E.coli* showed a reduced sensitivity against some antibiotics such as ticarcillin (18.5%), trimethoprim/sulfamethoxazole (14.8%), sulbenicillin (11.5%) and amoxicillin (11.1%). Since the number of other bacteria was too small, we will not discuss their resistance patterns to avoid misleading.
DISCUSSION

Infections of gastrointestinal tract may be localized and systemic, and a schematic representation is shown in figure 1. Guerrant et al. stated that the predominant microbes causing diarrhea in developing countries were enterotoxigenic *Escherichia coli* and rotavirus. Another study by Wasito and Bagus showed that in hospitalized children under 2 years old, *E. coli* (85.45%) was found to be the most frequent organism isolated, followed by *K. oxytoca* (9.09%), *Salmonella* sp (2.73%), *Vibrio cholerae* (1.82%), *Shigella* spp. (0.91%) and rotavirus (27.99%). It was interesting that in children under 2 years old many pathogenic bacteria such as *V. cholerae*, *Salmonella* and *Shigella* spp. were found in Wasito and Bagus’ study.

Furthermore, *Klebsiella oxytoca* that is a member of normal flora was found as one of etiological agent of diarrhea in hospitalized children under 2 years old. A study conducted by Omololu-Aso et al. found that five out of thirteen samples collected from patients (including a six month old baby) were positive for *Salmonella*. *Salmonella* isolated from the six month old baby was sensitive to amoxillin, cotrimoxazole, nitrofurantoin, gentamycin, ofloxacin and tetracycline, but was resistant to nalidixic acid and augmentin. The most prevalent *Salmonella* species found were *Salmonella typhi* and *Salmonella paratyphi*. There are many factors that may cause the presence of *Salmonella* in faeces, for example ingestion of contaminated food or drinking water.

**Figure 1.** Localized and systemic infections of gastrointestinal tract (modified from ref 7)

**Figure 2.** Resistance pattern of pathogenic *E. coli* against Amoxicillin, Sulbenicillin, Ticarcillin and Trimethoprim/Sulfamethoxazole 2005-2008
Our study revealed that the predominant bacteria that were isolated from the digestive tract were pathogenic *Escherichia coli*. However, we were only able to detect the pathogenic *E. coli* without determining the type of toxin produced, which was the limitation of this study. On the other hand, only one isolate of *Salmonella paratyphi* *A* was observed, while Rotavirus was not included in our study. Our results were different from another study conducted by Simadibrata et al, which found that chronic diarrhea causing microbes in adult were predominated by *C. albicans* (48.6%) and pathogenic *E. coli* (34.8%), while other bacteria represented low occurrences, i.e. *A. aerogenes* (3.6%), *M. tuberculosis* (3.6%), *K. oxytoca* (3.6%), *Salmonella paratyphi* (2.9%), *K. ozaenae* (2.2%), *K. pneumoniae* (2.2%), *A. dispers* (2.2%), *Geotrichum* (1.5%), *Pseudomonas spp.* (0.7%), *Y. enterocolitica* (0.7%), *C. perfingens* (0.7%), *S. sonnei* (0.7%) and *S. flexneri* (0.7%).

Olowe et al found that *E. coli* isolated from human clinical samples in Nigeria were resistant to tetracycline (91.6%), ampicillin (86.7%), sulphonamide (77.8%), cotrimoxazole (58.5%), cefuroxin (57.8%) and gentamicin (39.3%). More than 64% isolates were multi-drug resistant against 7 antibiotics tested. Sang et al studied the association of multi-drug resistant enteroaggregative *E. coli* with persistent diarrhea in Kenyan children, and they found that E. coli O44 was resistant to tetracycline, ampicillin, erythromycin, trimethoprim-sulphamethoxazole, and amoxicillin/clavulanate. Our study showed a slightly different resistance pattern, as we observed a reduced sensitivity of *E. coli* against some antibiotics such as ticarcillin, trimethoprim/sulfamethoxazole, sulbenicillin and amoxicillin.

In conclusion, in this study, gastrointestinal tract infection was mainly due to pathogenic *Escherichia coli*. This microbe showed reduce sensitivity against amoxicillin, sulbenicillin, ticarcillin, and trimethoprim/sulfamethoxazole.

REFERENCES


