Identification of pathogenesis pathway in basal-like breast cancer based on mutant p53 protein and topoisomerase-IIα expression

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Abstract

Background: Basal-like breast cancer is difficult to treat with standard regimen therapy, because it doesn’t express hormone receptors or epidermal growth factor receptors. Identification of oncogenesis pathway is expected to find molecules which can be used as target for therapy. One candidate molecule is topoisomerase-IIα whose expression is regulated by p53. This study aimed to compare the expression of mutant p53 proteins and topoisomerase IIα in basal-like and non basal-like breast cancer, and to determine the association between mutant p53 proteins and topoisomerase IIα in basal-like group.

Methods: The samples were 40 formalin fixed paraffin embedded tissues from verified triple negative breast cancer tissue. The samples were divided into 2 groups, basal-like and non basal-like breast cancer, based on cytokeratin - 5 (CK-5) expression. Mutant p53 proteins and topoisomerase IIα were stained using immunohistochemistry method, scored and compared. Statistical test used SPSS software version 16 for descriptive statistics, kappa test, normality test, comparison of two mean, and categorical comparison.

Results: Median (min-max) of mutant p53 protein expression in basal-like group was 21 (0-100), the non basal-like group was 2 (0-80), p = 0.061. Min-max expression of topoisomerase IIα in basal-like group was 263 (15-492), non basal-like group was 262 (0-481), p = 0.409. There was an association between mutant p53 positivity with breast cancer subtype (p = 0.027) and between mutant p53-topoisomerase IIα coexpression with breast cancer subtype (p = 0.018).

Conclusion: Co-expression of mutant p53 with topoisomerase IIα has the role in one of the pathway of basal-like breast cancer pathogenesis.

Keywords: basal-like breast cancer, mutant p53, topoisomerase-IIα
Breast cancer is the most common malignancy in women worldwide. In developing country the prevalence continues to increase and often diagnosed at an advanced stage. Breast cancer has heterogeneous clinical, pathological, histological, molecular, and response to treatment characteristics. In order to better understand and to find the appropriate treatment strategy for this disease, various attempts have been made to classify breast cancer.

Based on the expression of its mRNA, breast cancer is divided into 5 types: luminal A, luminal B, normal breast like, human epidermal growth factor receptor 2 (HER2) positive, and basal-like. Basal-like type has the most aggressive clinical behavior and poor prognosis. Basal-like breast cancer often do not express estrogen (ER), progesterone (PR) and HER2 receptors, hence triple negative breast cancer terminology. This type of breast cancer also frequently express high molecular weight cytokeratins, such as cytokeratin - 5 (CK-5). In other word, basal-like breast cancer is triple negative breast cancer that express high molecular weight cytokeratins. The non-expression of ER, PR, and HER2 in basal-like breast cancer is link to difficulties in therapy, because targeted therapy using standard hormone receptor with HER2 as its target, will not work. Therefore it is necessary to explain the characteristic of basal-like breast cancer, so that we will be able to explain the aggressiveness, and to find the potential molecule involved and hence design the right treatment.

In basal-like breast cancer, there are several known genetic defects, such as decreased expression of breast cancer genes 1 (BRCA1), genes responsible for repair of deoxyribose-nucleic acid (DNA) double stranded break. The high amount of DNA double stranded breaks in normal cells causes cell cycle stalled; but basal-like breast cancer seems to have high tolerance for BRCA1 defect, and cell cycle keep going on despite the high accumulation of DNA damage. This resistance is hypothetically caused by mutations in the p53 gene, a tumor suppressor gene. In a p53-gene-mutated-cell the cell cycle will not be interrupted even if there has been a significant defect in DNA.

One protein in which transcription is regulated by p53 is topoisomerase IIα. Topoisomerase IIα is an ubiquitous enzyme in the cell nucleus, which plays a role in the regulation of DNA topology. It also regulate many biological processes such as replication, segregation, transcription, regulation of chromatin structure and general gene expression.

These studies raised possibilities that pathogenesis of basal-like breast cancer could be mediated by pathway that involve the p53 gene and topoisomerase IIα, in which p53 tumor suppressor gene dysfunction causing increase expression of topoisomerase IIα.

Topoisomerase IIα is one of the molecular targets for chemotherapy, but the prognosis has not improved significantly. There are also several side effects caused by anti-topoisomerase IIα regimen such as secondary malignancies as well as cardiotoxicity. Therefore, ideally the regimen should only be given to cancer patients with positive topoisomerase IIα. It is important to study oncogenesis pathways of p53-topoisomerase IIα. If this pathway proves important in the pathogenesis of basal-like breast cancer then it could serve as the basis/justification for the treatment of breast cancer with a regimen targeting topoisomerase IIα.

**METHODS**

Ethical clearance was obtained from Institutional Review Board (IRB) of the Faculty of Medicine Universitas Indonesia. The study was conducted in the Immunopathology Laboratory Department of Anatomical Pathology Faculty of Medicine Universitas Indonesia, during January to December 2011. The study used 40 triple negative breast cancer tissues from formalin fixed paraffin embedded preparation. The samples were stained using automated immunostainer with three markers: CK-5, mutant p53, and topoisomerase IIα. For CK-5 (Rabbit anti CK-5, Biocare, dilution 1/100) and topoisomerase IIα (mouse anti topoisomerase IIα, DakoCytomation, dilution 1/500) we use machine-automated Benchmark XT immunostainer VENTANA with paraffin protocol. For staining of mutant p53 we used primary antibody mouse monoclonal anti-human p53 protein Clone DO-7 (DakoCytomation) 1:1500 dilution. Positive controls for each antibody were included using previously established positively stained tissue. The negative control from each case was also included.
Cell was categorized as positive for CK-5 if demonstrated cytoplasmic and membrane staining.\textsuperscript{14} The semi-quantitative assessments was as follows: negative if < 10\% of tumor cells show positive results, positive if \geq 10\% of tumor cells show positive results. Mutant p53 positive cell was nuclear stained cell and scored using proportion of 100 cells count, and categorized as positive if more than 10\% showed positive result, and negative if less than 10\%.\textsuperscript{15}

Topoisomerase II\(\alpha\) was positive if showed nuclear staining and scored by counting the positivity based on 500 cells count using 400x magnification.\textsuperscript{11} It was grouped as negative if < 25\% tumor cells show positive results, positive if \geq 25\% tumor cells show positive results.

The assessment of immunohistochemistry staining was done by two independent pathologists and the results were analyzed by Kappa test. Statistical test was using SPSS software version 16 for descriptive statistics, kappa test, normality test, comparison of two mean, and categorical comparison.

RESULTS

Forty two confirmed triple negative cases were obtained, among which 2 cases were excluded due to minimal amount of tumor tissue. Of the 40 cases, 20 cases were positive for CK-5 so it was grouped as the basal-like breast cancer, and the other 20 cases were negatively stained with CK-5 and were grouped as the non basal-like breast cancer. Assessment of mutant p53, topoisomerase II and CK-5 was performed by two independent pathologists, the kappa values were as followed: kappa values for mutant p53 = 0.866, p < 0.001; kappa values for topoisomerase II\(\alpha\) = 1, p = 0.002; kappa values for CK-5 = 0.879, p < 0.001; all showed good concordance.

Table 1. is about the median score of Topoisomerase II\(\alpha\) and mutant p53 expression in basal-like breast cancer and non basal-like breast cancer.

<table>
<thead>
<tr>
<th>Marker expression</th>
<th>Type of breast cancer</th>
<th>Basal-like n (%)</th>
<th>Non basal-like n (%)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Topoisomerase II(\alpha) median (min - max)</td>
<td>262 (0-481)</td>
<td>263 (15-492)</td>
<td>0.409</td>
<td></td>
</tr>
<tr>
<td>Mutant p53 median (min - max)</td>
<td>2 (0-80)</td>
<td>21 (0-100)</td>
<td>0.061</td>
<td></td>
</tr>
</tbody>
</table>

*Chi-square

Table 2. showed positivity of Topoisomerase II\(\alpha\) and mutant p53 in basal-like breast cancer and non basal-like breast cancer, in this table we can see there was an association between mutant p53 expression and the type of breast cancer, in the basal-like group mutant p53 expressed more often compared to non basal-like group.

Table 3. is about the correlation between expression of mutant p53 and Topoisomerase II\(\alpha\) in basal-like breast cancer, it showed that there was co-expression of mutant p53 and topoisomerase II\(\alpha\) in half of

Table 4. Proportion of p53 mutant-topoisomerase II\(\alpha\) co-expression in basal-like breast cancer and non basal-like breast cancer.

<table>
<thead>
<tr>
<th>Mutant p53</th>
<th>Topoisomerase II(\alpha) co-expression n (%)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Basal-like</td>
<td>10 (50%)</td>
<td>10 (50%)</td>
</tr>
<tr>
<td>Non basal-like</td>
<td>3 (15%)</td>
<td>17 (85%)</td>
</tr>
</tbody>
</table>

*Chi-square
the basal-like cases, but there was no significant difference between mutant p53 and topoisomerase IIα expression in the whole cases of basal-like breast cancer.

Table 4. showed proportion of p53 mutan-
Topoisomerase IIα co-expression in basal-like breast cancer and non basal-like breast cancer, in this table we can see there was significant difference in co-
expression mutant p53-topoisomerase IIα between basal-like breast cancer (50%) and the non basal-like breast cancer (15%).

Figure 1 showed the positivity of CK-5, mutant p53, and topoisomerase IIα, immunohistochemistry in breast cancer tissues.

**DISCUSSION**

In this study the positive expression of mutant p53 in basal-like breast cancer (65%) was higher than that of the non-basal-like (30%). This is in line with Conforti, et al. who found 51% positivity of mutant p53 in basal-like breast cancer, while in the non-basal like the positivity was only 11%. The explanation for this phenomenon was that as much as 80% - 90% basal-like breast cancer were associated with reduced expression of BRCA1. BRCA1 is a protein that has a role in various cellular processes such as response to DNA damage repair, control check points in the cell cycle, transcription modulation, and ubiquitination. In normal condition cells with reduced levels BRCA1, either due to mutation or promoter’s methylation should undergo cessation of the cell cycle and eventually death. In basal-like breast cancer it seems that the tumor cells are not responding to the lack of BRCA1 levels so that the cell cycle continue. This phenomenon might be caused by mutation of the tumor suppressor gene p53, allowing genetically defective cells to continue replicating and eventually transforming into cancer cells. This explanation is consistent with the results of the study by Holstege, et al. who found that all breast cancers with BRCA1 expression defects were also contained p53 mutations, while other studies found 60%-77% p53 mutations in breast carcinoma with BRCA1 defects.

No difference was found in topoisomerase IIα expression between basal-like breast cancer and the non basal-like, although the basal-like group has slightly higher positivity (75%), compared to non basal-like (60%). This finding is actually consistent with the results reported by Romero, et al. They stated that topoisomerase IIα protein expression which was measured by immunohistochemistry method, was higher in the basal-like breast carcinoma (72.72%), compared with the other subtypes: luminal type B (57.14%), HER2 positive (62.5%), luminal A (9%), and normal-like (0%).

This study also assessed the relationship between mutant p53 expressions with topoisomerase IIα expression in basal-like breast cancer; although we didn’t find any significant association, but it showed that mutant p53-topoisomerase IIα co-expressed in half of cases. This didn’t explain the hypothesis of mutant p53 and topoisomerase IIα role in basal-like breast canacer. There might be other carcinogenesis pathway involved in the pathogenesis of basal-like breast cancer. They were: epidermal growth factor receptor (EGFR), proto-oncogene c-kit (c-KIT), and BRCA1. Another possibility was that the biological trait of topoisomerase IIα doesn’t expressed continuously in the cell cycle. The highest expression is in late S phase and peaked in growth2 - mitosis (G2-M) phase. Because of this feature topoisomerase IIα considered as cell proliferation surrogate marker, and doesn’t expressed in the cell constantly, instead it depends on the cell cycle status.
The non association between mutant p53 and topoisomerase IIα may be caused by the method used to assess p53 status. In this study we used immunohistochemistry to detect the presence of mutant p53, because mutant p53 have longer half-life than the wild type variant, according to literature. Half-life of wild-type p53 is very short and it's basically expressed in a very small amount below the detection threshold of immunohistochemistry. \(^{22,23}\) Sjogren, et al\(^\text{22}\) stated that mutant p53 detection using immunohistochemistry produce false positive results as much as 30% and false-negative results around 33% when compared to measurements using copy dna (cDNA) sequencing. It was mentioned that increased expression of p53 by immunohistochemistry standard methods can represent mutations in the p53; verification using cDNA sequencing are considered as gold standard.\(^{22}\) False positive staining by immunohistochemistry method can be due to cellular stress induced p53 wild-type stabilization. Whereas the incidence of false negatives may occur due to mutation causing stop codons, deletions, or mutation causing destabilization of the protein.\(^{22}\) Truncation at the carboxyl end of p53 could cause false negative result, since the truncated mutant p53 protein will be degraded due to lost of several important function, for example the function for DNA binding domain, nuclear localization signals, and oligomerization domain\(^{24}\).

We also assess the relationship between type of breast carcinoma with the incident of topoisomerase IIα-mutant p53 co-expression, and it revealed significant results with \(p = 0.018\). Half of basal-like breast cancer showed topoisomerase IIα-mutant p53 co-expression, which is higher than the co-expression in non basal-like group which only expressed in 3 samples (15%). This supports the hypothesis of mutant p53 and topoisomerase IIα involvement in basal-like breast cancer pathogenesis. According to Wang, et al\(^\text{7}\) and Sandri, et al\(^\text{6}\) p53 has the ability to regulate topoisomerase IIα expression, in \textit{in-vitro} studies they found that p53 has the ability to regulate topoisomerase IIα transcription, by interacting with topoisomerase promoter (-32 to +90) in dna sequence cytosine cytosine adenine adenine tyrosine (CCAAT) segments. Wild-type p53 can act as a trans-acting elements capable of suppressing transcription of topoisomerase IIα, while mutant p53 has less ability to suppressed topoisomerase IIα expression. This finding can also serve as the consideration for treatment planning using anti topoisomerase IIα drug in part of basal like breast cancer.

In conclusion, this study showed a significant association between mutant p53 expression and basal like subtype. There was no association between p53 and topoisomerase IIα expression in basal like group. Co-expression of mutant p53-topoisomerase IIα occurred significantly more often in basal like, which indicates the role of mutant p53-topoisomerase IIα pathway in basal like pathogenesis in part of cases.

**Conflicts of interest**

The authors affirm no conflict of interest in this study.

**Acknowledgments**

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**REFERENCES**


