

## Cytogenetic Pattern of Acute Lymphoblastic Leukemia (ALL) in the Yogyakarta and Central Java Province, Indonesia

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### ABSTRAK

Penelitian ini bertujuan untuk melihat gambaran sitogenetika penderita ALL (Acute Lymphoblastic Leukemia). Gambaran sitogenetika ini penting untuk menentukan penatalaksanaan penderita tersebut. Empat belas pasien ALL yang berobat ke rumah sakit di Yogyakarta dan Semarang sejak Juni 1992 sampai November 1992 telah diperiksa gambaran sitogenetikanya dengan metode G-banding. Hasil pemeriksaan menunjukkan 4 penderita dengan kelainan struktural kromosom berupa t(9;22) (q34.1;q11.21) pada 2 pasien, t(4;11) (q21;q23) pada 1 pasien, del 11, p13-14 pada 1 pasien; 9 penderita dengan kelainan jumlah kromosom berupa hiperploidi, poliploidi, tetraploidi, atau monosomi, 3 diantaranya termasuk kelainan kromosom ganda (struktural dan numerikal); 4 penderita tidak ditemukan kelainan kromosom. Pada penderita dengan hanya hiperploidi > 50 kromosom dan gambaran sitogenetika normal setelah pengobatan menunjukkan remisi/remisi parsialis. Sedangkan 3 penderita dengan kelainan kromosom ganda menunjukkan perjalanan penyakit lebih progresif. Disimpulkan bahwa kelainan struktural pada ALL mempunyai arti penting dalam menentukan penggolongan dan prognosis penyakit.

### ABSTRACT

The aim of this investigation is to analyse the cytogenetics of ALL patients. Cytogenetic analysis is important in determining the management of those patients. Fourteen ALL patients, hospitalised in several hospitals in Yogyakarta and Semarang since June 1992 to November 1992, were studied by G-banding method. The results showed 4 patients with structural abnormality t(9;22) (q34.1;q11.21) (2 patients), t(4;11) (q21;q23) (1 patient), and del 11, p13-p14 (1 patient); 9 patient with numerical abnormality such as hyperploidy, polyploidy, tetraploidy and monosomy, three of them showed double abnormalities (structural and numerical); 4 patients with normal karyotype. Patients with previously have hyperploidy > 50 chromosom and normal karyotype, after treatment showed remission/partial remission. Patients with double abnormalities were clinically more progressive. It is concluded that structural chromosomal abnormality in ALL is important to determine the group and prognosis of the disease.

**Key words :** ALL - cytogenetics - translocation - deletion - hyperploidy - classification - prognosis.

### INTRODUCTION

Cancer is the result of accumulation of multiple genetic changes.<sup>1</sup> Each alteration, whether an initiating or a progression event, may be mediated through a gross chromosomal change and therefore has the potential to be cytogenetically visible.<sup>2</sup>

The common tumor chromosomal aberrations are classified as structural or numeric. Structural alterations include translocations, inversions, deletions,

insertions and amplifications, whereas numerical abnormalities are losses or duplication of whole chromosomes.<sup>3</sup> In hematological malignancy such as leukemia, analysis of the chromosomes also has been extensively studied, which revealed that almost all leukemia carry a karyotype abnormality.

Hematological malignancy particularly leukemia is still a problem in Indonesia. Cases recorded at Dr. Sardjito Hospital in the Yogyakarta province from 1987 to 1990 included Hodgkin lymphoma (6.29%),

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non Hodgkin lymphoma (46.85%), multiple myeloma (3.14%), lymphoid leukemia (21.86%), myeloid leukemia (19.86%) and other types of leukemia (1.98%).<sup>4</sup> The frequency of malignancy cases from year to year was increasing. In 1987 there were 884 cases, in 1988 1164 cases, in 1989 1190 and in 1990 1282 cases. The total number of malignancy cases at Dr. Sardjito Hospital Yogyakarta were 4520 cases.<sup>4,5</sup> In Semarang, Sunjoyo reported that there were 84 cases of hematological malignancies between 1984-1986, included non Hodgkin lymphoma (42.9%), myeloid leukemia (32.4%), lymphoblastic leukemia (17.2%), multiple myeloma (3.7%) and Hodgkin lymphoma (2.3%).<sup>6</sup>

The diagnosis of leukemia in Yogyakarta or Indonesia in general is based on the morphological finding of blood lineage cells, in blood or bone marrow smear stained with routine staining such as Giemsa, Wright, May Grunwald, and examined under regular light microscope (FAB Classification).<sup>5</sup>

Therapy using standard regimen therapy showed no satisfactory results;<sup>5</sup> this may be due to the timing of patients arrival at the hospital. Usually patients came to the hospital at the terminal state of the disease or might be due to inaccuracy of diagnosis. In 1988 a group of hematologist proposed a new classification of leukemia based on morphology, immunophenotyping and cytogenetics (MIC).<sup>7</sup> Classification which merely based on morphology could not classify the type of leukemia.<sup>7</sup> Using cytogenetic analysis will increase the accuracy of the diagnosis, so that the therapy can be determined and the prognosis can be predicted.<sup>8,9,10</sup> In Indonesia, the diagnosis of leukemia using cytogenetic analysis has not been routinely used.

The aim of this study is to analyse the cytogenetic pattern of ALL patients in Yogyakarta and Central Java, Indonesia, in the hope that therapy could be correctly given to the patient and prognosis of the disease can be predicted.

## MATERIALS AND METHODS

### Samples

One ml of heparinized peripheral blood and 0.5 ml of bone marrow aspirate from patients diagnosed as leukemia at Dr. Sardjito Hospital (Yogyakarta), Kariadi Hospital (Semarang), and Telogorejo Hospital (Semarang).

### Methods

**Arresting :** Colcemid was used in this investigation as arresting agent to stop mitosis at metaphase.<sup>3</sup> Colce-

mid is a mitotic spindle inhibitor, which can block the formation of spindle fibres. Normally, spindle fiber was attached to the centromere of each chromosome before mitotic division and pull the chromatids apart by contraction at anaphase.<sup>3</sup>

**Hypotonic Shock :** The hypotonic solution used was 0.075M KCL since it has been known, that this solution is the least damaging to chromosomes substructure. A hypotonic solution is a solution with a lower salt concentration than that of the cytoplasm of cells, so there will be a movement of H<sub>2</sub>O from the hypotonic solution into the cell.<sup>3</sup>

**Fixation :** The fixative solution was freshly made of 3 parts of absolute methanol to 1 part of glacial acetic acid. The volatile alcohol plays a role in flattening and spreading the chromosomes during air drying. Glacial acetic acid will rupture the membrane of the cell.

**Slide Preparation and Staining :** Slides were washed in distilled water, dried at room temperature and stained with Giemsa.<sup>11</sup>

### Giemsa Staining and Banding

Giemsa staining is a method to stain the chromosomes. Giemsa stain is a Romanowsky-type dye mixture. Azure B and eosin in the Giemsa bound to DNA.<sup>11</sup> The solutions needed are :

1. Giemsa stain concentrate, made by adding 1.0 g Giemsa powder to 66 ml methanol and 66 ml glycerin, stirred for 2 days at room temperature, prepared at least 2 weeks before use. Stored in a dark container in a refrigerator.
2. Phosphate buffer (pH 6.8), which consists of 0.025M KH<sub>2</sub>PO<sub>4</sub> (3.4 g/l), and the pH was adjusted using 50% NaOH.

Giemsa staining solution was prepared by mixing 5 ml of Giemsa stain concentrate and 45 ml of phosphate buffer (pH 6.8).

Giemsa banding (G banding) is a method to obtain bands in chromosomes by treating slides with a protease such as trypsin or incubate the slides in hot salinecitrate. The chromosome banding pattern obtained reflects both structural and functional composition of the chromosomes.<sup>10,11</sup>

Solutions needed are :

1. Giemsa stain concentrate
2. Phosphate buffer (pH 6.8)
3. Trypsin-EDTA 10X concentrate (trypsin (1:250) 5 g/l and EDTA 2 g/l).

Working staining solution is prepared by mixing 26 ml buffer (pH 6.8), 7 ml methanol, 0.6-1.0 ml 10X trypsin EDTA and 0.8 ml Giemsa stain concentrate.

### Chromosome Analysis

Chromosomes are prepared by direct bone marrow and peripheral blood culture developed in our laboratory. Marrow cells were cultured in RPMI-1640 medium supplemented with 15% fetal calf serum. Only  $\pm$  0.1 ml of sedimented cells were processed from each centrifuge tube. The cells were exposed to colcemid (0.06 mg/ml) for 25 minutes and then to hypotonic solution i.e. KCl (0.075 mol/l) for a total of 32 minutes (including mixing, standing and centrifuge time). Slides were fixed in 3:1 methanol acetic acid (v/v) and dried on a hot plate. Slides were subjected to Giemsa staining and banding.

For each case, 10-20 metaphases (mean 15) were studied by direct microscopy to determine the modal number and to identify the malignant stem line. Karyotype were prepared from photographic prints. Chromosomal abnormalities were classified according to the International Conventions.<sup>10,11,12</sup> The definition of an abnormal stem line was defined according to that proposed by the Second International workshop on chromosome in leukemia. A case was considered abnormal if it had an abnormal clone, regardless of the proportion of normal metaphase.<sup>13</sup>

### RESULTS

The patients were diagnosed when they were hospitalised. There were 14 patients analysed cytogenetically. Out of those 14 patients 8 were males and 6 were females (Table 1). The range of the age was between 2-20 years old. There were 4 patients under 5, 7 patients between 5 and 12, and 3 patients more than 12 years old (Table 2). The diagnosis was based on FAB classification (morphology) by Wright or Giemsa staining.

Table 1. Patients with ALL

Sex	n	%
Male	8	57.1
Female	6	42.9
Total	14	100

n = number of patient

The cytogenetic analysis on peripheral blood and bone marrow samples were carried out on at least 10 metaphases from one slide. The findings were as follows (Table 2) :

- 1 patient of 46 XY, t(4;11) (q21;q23) from peripheral blood and 53 XXY from bone marrow.
- 1 patient of 46 XY t(9;22) (q34.1;q11.21) (Philadelphia chromosome positive) from peripheral blood and 56 XY from bone marrow.
- 1 patient of t(9;22) (q34.1;q11.21) from peripheral blood, Ph+ from bone marrow.
- 7 patients of abnormality in chromosome number from bone marrow i.e. 52 XY mozaic, 91 XX polyploidy mozaic, hyperploidy > 50 chromosomes mozaic, 87 XY polyploidy mozaic (25%), 92 XXYY tetraploidy mozaic, and monosomies 45 XX-7 and 45 XY-17.
- 4 patients showed no structural and numerical chromosome abnormality.

Table 2. Age, Sex, Cytogenetic finding of ALL

No.of patient	Age (years)	Sex	Cytogenetic finding	Source
1.	4	M	46 XY, mozaic, 52 XY	BM
2.	4	F	46 XX, mozaic, 91 XX polyploidy	BM
3.	5	F	46 XX	
4.	6	M	46 XY t(4;11)(q21,q23)/53 XXY (4+1;5+1;6+1;11+1;16+1;21+1;X+1)	P/BM
5.	5	F	45 XX-7	BM
6.	3	M	46 XY t(9;22)(q34.1,q11.21)(Ph+)/ 56 XY hyperploidy (5+1)(6+1)(8-1)(13+1)(14+1)(16+1)(17-1)(18+2)(21+3)(22+2)	P/BM
7.	20	M	46 XY, hyperploidy > 50 Chromosome	BM
8.	6	M	46 XY	
9.	11	M	46 XY, 87 XY polyploidy 25%	BM
10.	4	F	46 XY t(9;22)(q34.1,q11.21)/Ph+	P/BM
11.	9	F	46 XX	
12.	6.5	F	46 XX	
13.	20	M	46 XX, 92 XXYY tetraploidy	BM
14.	19	M	45 XY-17 del 11,p13-p14	BM

BM = source of karyotyping analysis is from bone marrow

P/BM = source of karyotyping analysis is from bone marrow and peripheral blood

M = male

F = female





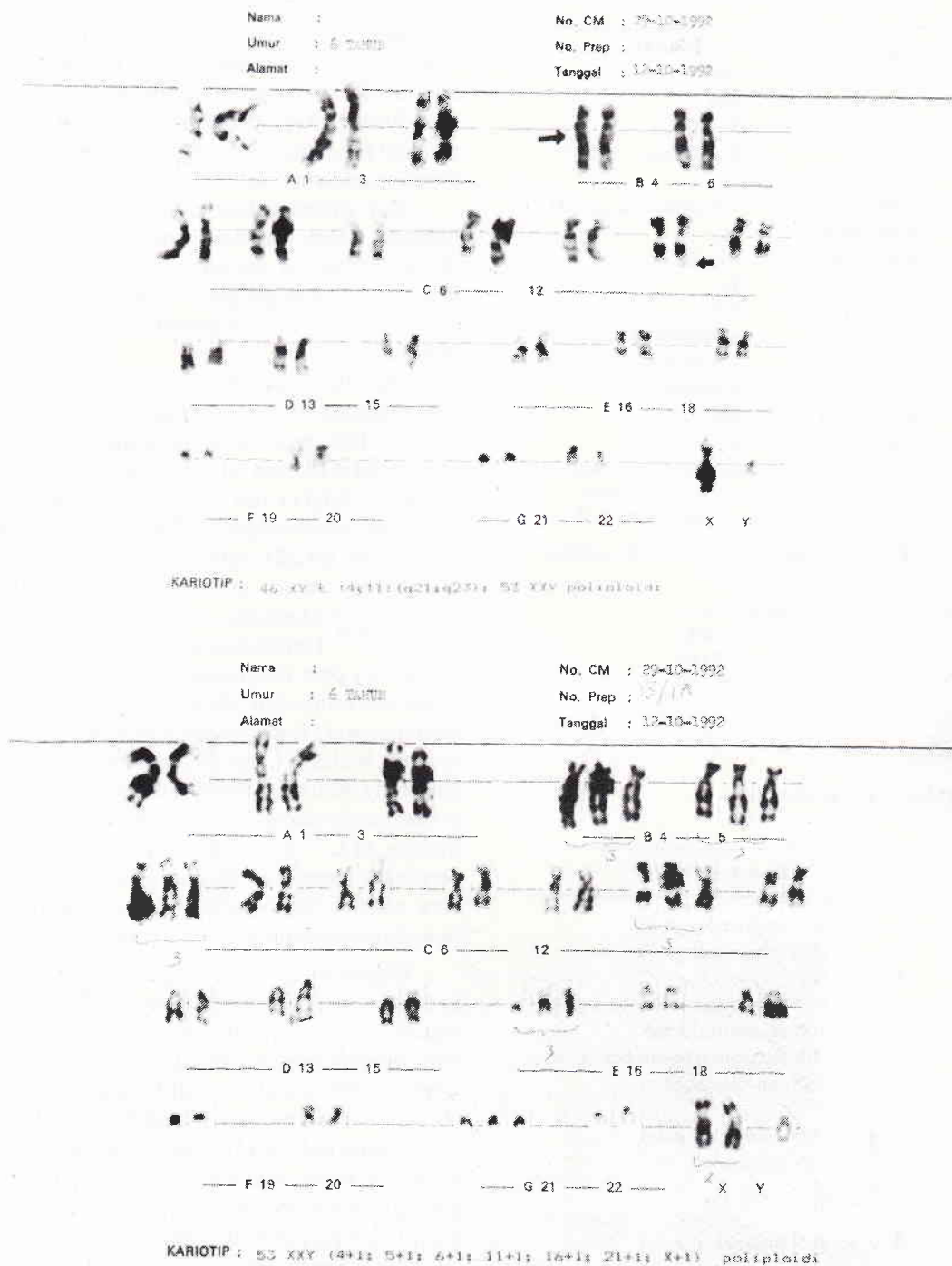


Figure 1. Karyotype of an ALL-L1 patient showing  
 a) 46 XY, t(4;11)(q21;q23) and  
 b) 53 XXY from bone marrow culture  
 Karyotyping was done from G-banded chromosome  
 t = translocation q = long arm of the chromosome  
 G = Giemsa  
 ALL-L1 = Acute lymphoblastic leukemia type L-1

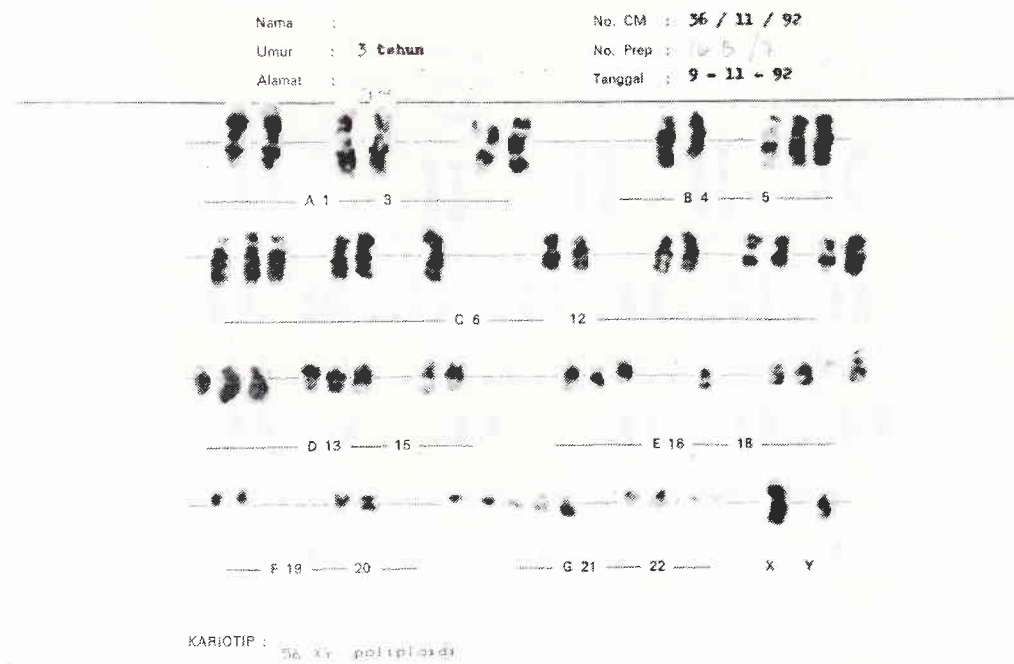
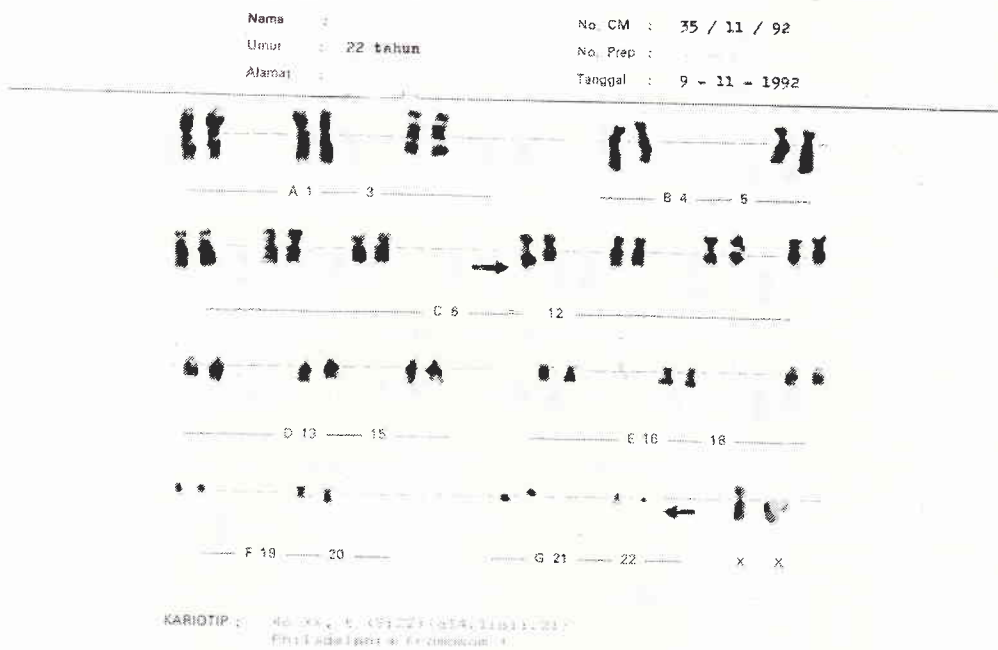


Figure 2. Karyotype of an ALL-L1 patient showing  
 a) 46 XY, t(9;22)(q34;q11.21) (Ph+) and  
 b) 56 XY (poliploidy)  
 Karyotyping was done from G-banded chromosome  
 t = translocation q = long arm of the chromosome  
 (Ph+) = Philadelphia chromosome positive  
 G = Giemsa  
 ALL-L1 = Acute lymphoblastic leukemia type L-1

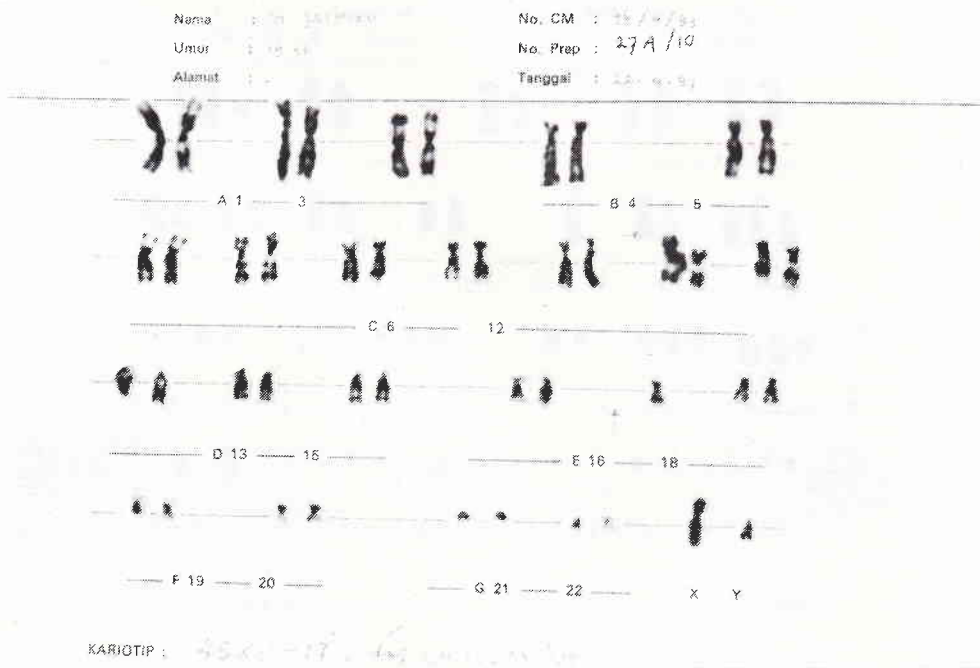
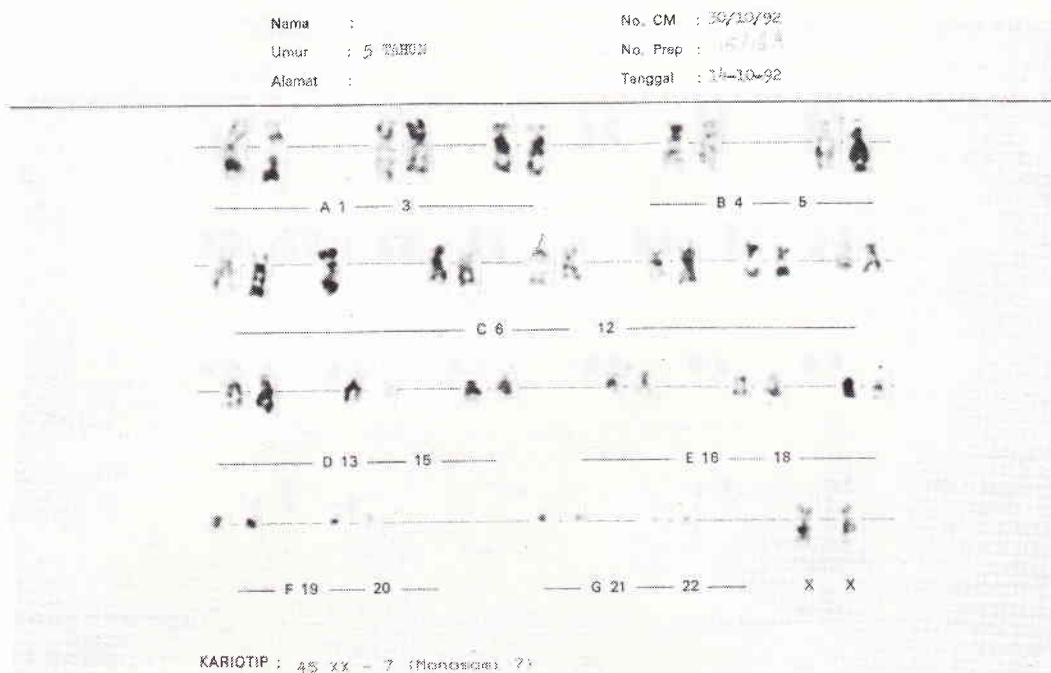


Figure 3. Karyotyping of two ALL-L1 patient showing  
 a) monosomy 46 XX-7 and  
 b) monosomy 45 XY-17  
 karyotyping was done from G-banded chromosome  
 G = Giemsa  
 ALL-L1 = Acute lymphoblastic leukemia type L-1

## CONCLUSION

Our results suggested that multiple abnormalities and monosomies indicate poorer prognosis compare to hyperploidy, poliploidy and normal karyotype.

Patients with translocation or deletion of whole chromosome should be categorized as high risk patients regardless the Hb, WBC and platelets count.

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