Decreased sensitivity of several anticancer drugs in TMEPAI knockout triple-negative breast cancer cells

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ABSTRACT

BACKGROUND Transmembrane prostate androgen-induced protein (TMEPAI) was reported to be highly amplified in the majority of patients with triple-negative breast cancer (TNBC). TMEPAI is related to poorer prognosis, limited treatment options, and prone to drug resistance compared with other proteins. One of the established markers to determine cancer resistance to drugs is the increased expression levels of drug efflux transporters. However, the role of TMEPAI in cancer resistance to drugs has not been elucidated. This study was aimed to investigate whether TMEPAI participates in cancer resistance to drugs by regulating drug efflux transporters.

METHODS TMEPAI knockout (KO) cells were previously developed from a TNBC cell line, Hs578T (wild-type/WT), using a CRISPR-Cas9 system. The expression levels of drug efflux transporters were determined in Hs578T-KO and Hs578-WT by quantitative reverse transcriptase polymerase chain reaction. Cytotoxic concentration 50% (CC50) of several anticancer drugs (doxorubicin, paclitaxel, and cisplatin) were determined in the two cell lines via 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium assay.

RESULTS The results showed that the mRNA expression of P-glycoprotein (P-gp) and breast cancer resistance protein (BCRP) was significantly increased in Hs578T-KO compared with that in Hs578-WT cells. CC50 of several anticancer drugs (doxorubicin, cisplatin, and paclitaxel) were determined in the two cell lines via 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium assay.

CONCLUSIONS TMEPAI participated in the regulation of mRNA expression levels in drug efflux transporters (P-gp, BCRP, and multidrug resistance-associated protein 1). Further studies are necessary to confirm whether this finding might be dependent on the development of cancer cell sensitivity to anticancer agents.

KEYWORDS BCRP, MRP1, P-glycoprotein, TMEPAI, triple-negative breast cancer cell
Transmembrane prostate androgen-induced protein (TMEPAI) was first reported as a novel androgen-regulated gene. TMEPAI is also known as a prostate transmembrane protein, androgen induced 1, and solid tumor-associated gene 1. It is related to tumor relapse in the human prostate cancer xenograft model that is strongly androgen-dependent. Instead of androgen, TMEPAI is induced by epidermal growth factor in breast and ovarian cancer cell lines and in breast primary tumors and transforming growth factor-β (TGF-β) in many cancer cell lines. In patients with TNBC, TMEPAI was reported to be highly amplified in 68.8% of patients and correlated with short survival time. Previous analysis of TMEPAI’s prognostic significance suggested that high TMEPAI expression is correlated with poor prognosis with a hazard ratio of 1.88 (p < 0.05).

Previous study of Wardhani et al. showed that TMEPAI also plays a role in cell proliferation. TMEPAI was also reported as a converter for TGF-β from a tumor suppressor in the early stages of carcinogenesis to a tumor promoter in advanced stages of carcinogenesis because of the inhibition of the TGF-β canonical Smad pathway and the enhancement of the TGF-β1-induced PI3K/AKT signaling pathway. Nevertheless, TMEPAI has been shown to contribute to TGF-β1-induced epithelial-mesenchymal transition (EMT) in lung cancer cells, and it further induces drug resistance in pancreatic and lung cancer. However, to date, the role of TMEPAI in cancer resistance to drugs has not been elucidated. This study was aimed to investigate whether TMEPAI might participate in drug resistance by regulating drug efflux transporters.

**METHODS**

**Cell culture**

This in vitro experimental study involved a TNBC cell line, Hs578T. The TNBC cell line Hs578T was obtained from Tsukuba University. Hs578T TMEPAI knockout (KO) cells were previously established from Hs578T wild-type (WT) using a CRISPR-Cas9 system. Both WT and KO Hs578T were cultured in Dulbecco’s modified essential medium (Invitrogen) supplemented with 10% fetal bovine serum (Gibco), 10 μg/ml insulin, 100 units/ml penicillin G, and 0.1 mg/ml streptomycin sulfate (Wako). Cells were maintained in a 5% CO₂ incubator at 37°C. Cells were trypsinized at a confluency of about 80–90% and subjected to RNA isolation and quantitative reverse transcriptase polymerase chain reaction (qRT-PCR) analysis.

**qRT-PCR**

We isolated total RNA by using a Total RNA Mini Kit (Geneaid) from cultured cells, and RNA was converted to cDNA by using Transcriptor First Strand cDNA Synthesis Kit (Roche). RT-PCR was performed using FastStart Essential DNA Green (Roche). Primers used were purchased from Integrated DNA Technologies (IDT, Singapore) for β-actin, P-gp, MRP1, and BCRP. The sequences for P-gp were Fwd: 5'-TTACATTCAAGTCTTCACTTGGT-G-3'; P-gp Rev: 5'-TCTTGTCGGATTATAGCATGA-3'; the sequences for BCRP were Fwd 5'-TTCTGCTTCATTATAGCATGA-3'; BCRP Rev: 5'-TCCCCCTCCTGGACTCCATGC-3'; and the sequences for MRP1 were Fwd 5'-ATGTCACGGAATACCCAG-3' and Rev: 5'-GAAACTGAACCCCTCCCTCC-3'. β-actin was used as a housekeeping gene with the following sequences: β-actin Fwd: 5'-GCTGGAAGGTGGACAGCAG-3', β-actin Rev: 5'-GGCATCGTGATGGACTCCG-3'.

Approximately 20 μl of the reaction was used in a 32-well plate on qRT-PCR Light Cycler Nano (Roche, USA). RT-PCR conditions comprised a three-step amplification process (45 cycles) in a specific annealing temperature of 52°C for P-gp and BCRP but 53°C for MRP1. The quantification cycle was analyzed by relative quantification using Livak method and described as normalized expression ratios of P-gp, BCRP, and MRP1 to β-actin.

**Cytotoxic concentration**

Cytotoxic concentration was measured based on cell viability via 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (or MTS) assay by following the protocol in CellTiter 96® Aqueous One Solution Cell Proliferation Assay. About 1 × 10⁴ cells were seeded in a 96-microplate, and cells were grown until about 70% confluency. Subsequently, anticancer drugs were added 24 hours before performing MTS assay with cell confluence of at least 70%. Doxorubicin, paclitaxel, and cisplatin were dissolved in dimethyl sulfoxide (DMSO) and diluted until the required concentrations were obtained. DMSO used in final concentrations did not exceed 0.01%. DMSO was
used as control in this experiment. Concentrations used for doxorubicin, paclitaxel, and cisplatin were from 0.5–50 µM. CC50 of the three drugs was calculated from the extrapolation from log-linear equations.

**Data analysis**

The differences between the two groups, Hs578T WT and Hs578T KO, was analyzed using Student’s t-test. Significance was determined at $p < 0.05$.

**RESULTS**

**Cell morphology**

The morphology of Hs578T WT and Hs578T KO cells is shown in Figure 1, a and b. No marked difference was observed in WT and KO cells.

![Figure 1](image1.png)

**Figure 1.** Morphology in KO cells was not remarkably different from that in WT cells: (a) TNBC cell line, Hs578T, which has TMEPAI WT (Hs578T WT); (b) TNBC cell line, Hs578T, which has TMEPAI KO (Hs578T KO) by CRISPR-Cas9 systems. KO=knockout; WT=wild-type; TNBC=triple-negative breast cancer; TMEPAI=transmembrane prostate androgen-induced protein

**TMEPAI modulates mRNA expression of drug efflux transporters in Hs578KO cells**

Our results showed that TMEPAI affected the drug efflux transporters, especially P-gp and BCRP (Figure 2). A slight increase in MRP1 was also found.

![Figure 2](image2.png)

**Figure 2.** TMEPAI changed the mRNA expression level of drug-efflux transporter: (a) P-gp mRNA expression level in Hs578T KO significantly increased compared with that in Hs578T-WT; (b) BCRP mRNA expression level in Hs578T KO significantly increased compared with that in Hs578T-WT; (c) MRP1 mRNA expression level increased in Hs578T-KO compared with that in Hs578T-WT. The mRNA expression was calculated as relative expression levels after normalization by β-actin as a housekeeping gene. All experiments were conducted in duplicate and repeated three times. Statistical analyses were performed using independent t-test. *$p < 0.05$. TMEPAI=transmembrane prostate androgen-induced protein; P-gp=P-glycoprotein; KO=knockout; WT=wild-type; BCRP=breast cancer resistance protein; MRP1=multidrug resistance-associated protein 1
Decreased sensitivity to anticancer drugs in Hs578T KO cells

In all the three most common anticancer drugs used in TNBC, a significant decrease in sensitivity to several anticancer drugs was noted (Figure 3). The increased sensitivity was most obvious in doxorubicin (2.8 times) and paclitaxel (2.3 times).

![Doxorubicin sensitivity graph](image1)

![Paclitaxel sensitivity graph](image2)

![Cisplatin sensitivity graph](image3)

**Figure 3.** Decreased anticancer sensitivity in TMEPAI KO TNBC. Hs578T WT and KO were analyzed by MTS assay after 24 hours of anticancer treatment. All experiments were conducted in triplicate. Statistical analyses were performed using independent t-test. *p < 0.05. CC50=cytotoxic concentration 50; TMEPAI=transmembrane prostate androgen-induced protein; KO=knockout; TNBC=triple-negative breast cancer; WT=wild-type

**DISCUSSION**

Given the high expression of TMEPAI in most patients with TNBC and its correlation with short survival and poor prognosis, we were interested to investigate it further. Regarding the molecular mechanism of TMEPAI, Watanabe et al. and Singha et al. reported that TMEPAI has tumorigenic activity in lung cancer and TNBC. The high expression of TMEPAI is also associated with oncogenic Snail expression. Therefore, these reports suggested that TMEPAI may be correlated with tumorigenesis and drug resistance. However, the molecular mechanisms of TMEPAI in tumorigenesis and drug resistance remain unclear. This paper was focused on anticancer drug resistance.

In a preclinical study, Lehmann et al. identified that human TNBC subtypes display gene expression patterns. Hs578T was reported as a mesenchymal-like subtype suggested for preclinical studies on the TGF-β pathway. This result was in accordance with TMEPAI research because of its involvement in the TGF-β pathway. We then considered morphological alterations in drug resistance. Chen et al. reported morphological changes in doxorubicin-resistant cells compared with non-resistance cells using the BT20 cell line, an unclassified TNBC. They confirmed alterations in EMT markers, Snail and Twist, and morphological changes. In this model, no remarkable differences was observed in cell morphology between KO and WT (Figure 1, a and b). Singha et al. reported that knockdown of TMEPAI TNBC cells demonstrates higher expression of Snail compared with WT. However, further experiments are necessary to determine whether the upregulation of EMT markers takes place in this models.

Currently, cytotoxic drugs are the backbone of TNBC treatments. Initially, the drugs show good efficacy, but cancer cells develop resistance to anticancer agents after a short period. Doxorubicin, paclitaxel, and cisplatin are commonly used in chemotherapy regimen. Doxorubicin kills cancer cells by intercalating into DNA and disrupting topoisomerase II-mediated DNA repair. Moreover, doxorubicin generates free radicals to harm cellular membranes. Paclitaxel is known to induce mitotic arrest, which leads to cell death. By contrast, cisplatin works by crosslinking with purine bases on DNA, interfering with the DNA repair mechanism, causing DNA damage, and promoting apoptosis in
cancer cells. These anticancer drugs, which are used in TNBC, address DNA damage and cell cycles.

The overexpression of drug efflux transporters such as P-gp, BCRP, and MRP1 is one of the most common mechanisms in drug resistance. Doxorubicin, paclitaxel, and cisplatin are substrates for the three transporters, so increasing their expression will reduce the drug concentration in cancer cells. This finding is in accordance with this results. The increased expression of P-gp, BCRP, and MRP1 might lead to the efflux of anticancer agents from the cells. Hence, Hs578T KO demonstrated higher CC50 than WT cells.

This study investigated whether TMEPAI participates in drug resistance via the regulation of drug efflux transporters. This study results showed an enhancement in drug efflux transporters, followed by decreasing sensitivity of some anticancer agents. In line with this results, a previous study by Singha et al showed that TMEPAI inhibits TGF-β growth suppression and enhances PI3K/Akt signaling in TNBC. In addition, Bholi et al reported that inhibition of the TGF-β pathways prevents drug resistance in cancer stem-like cells from TNBC. Li et al also showed that activating the PI3K/Akt pathways promotes multidrug resistance for doxorubicin, paclitaxel, and cisplatin in cervical cancer cell line with increasing drug efflux transporter. Thus, activation of PI3K/Akt might be the underlying pathway responsible for the contribution of TMEPAI in drug resistance. However further studies need to verify this proposed pathway.

This study showed that the transporters were modulated in the TMEPAI KO cell line (Hs578T KO). The results demonstrated significant alterations in the mRNA expression levels of P-gp and BCRP after TMEPAI knocked out in the TNBC cell line (Figure 2, a and b). In accordance with these results, Hs578T KO showed decreasing sensitivity to anticancer drugs (CC50 increased; Figure 3). The clinical relevance and various expression levels of TMEPAI in patients with TNBC and cell lines were proven by Singha et al. They suggested that TMEPAI is an appropriate biomarker in distinguishing the staged development of TNBC. To complete those results, data on anticancer sensitivity were provided. TMEPAI is suitable for biomolecular targets to increase the sensitivity of patients with TNBC to anticancer drugs. Further experiments are warranted to elucidate the molecular mechanism of TMEPAI in anticancer sensitivity.

In conclusion, this results suggested that TMEPAI played a key role in the regulation of drug efflux transporters, particularly P-gp and BCRP. However, further mechanism-based studies are necessary to confirm this findings.

Conflict of Interest
Melva Louisa and Rianto Setiabudy are editorial board members but were not involved in the review or decision for the article.

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